



*Journal of Fish Biology* (2014) **85**, 151–188 doi:10.1111/jfb.12432, available online at wileyonlinelibrary.com

# **REVIEW PAPER**

# Early environment influences later performance in fishes

B. Jonsson\* and N. Jonsson

Norwegian Institute for Nature Research, Gaustadalléen 21, N-0349 Oslo, Norway

(Received 12 January 2014, Accepted 28 April 2014)

Conditions fish encounter during embryogenesis and early life history can leave lasting effects not only on morphology, but also on growth rate, life-history and behavioural traits. The ecology of off-spring can be affected by conditions experienced by their parents and mother in particular. This review summarizes such early impacts and their ecological influences for a variety of teleost species, but with special reference to salmonids. Growth and adult body size, sex ratio, egg size, lifespan and tendency to migrate can all be affected by early influences. Mechanisms behind such phenotypically plastic impacts are not well known, but epigenetic change appears to be one central mechanism. The thermal regime during development and incubation is particularly important, but also early food consumption and intraspecific density can all be responsible for later life-history variation. For behavioural traits, early experiences with effects on brain, sensory development and cognition appear essential. This may also influence boldness and other social behaviours such as mate choice. At the end of the review, several issues and questions for future studies are given.

© 2014 The Fisheries Society of the British Isles

Key words: epigenetic effects; growth; life-history traits; maternal effects; sex ratio; social behaviour.

# INTRODUCTION

Animal traits are influenced by early life experiences. For almost a century, research has revealed how environments of embryos influence skeleton and muscle development of ectothermic animals including fishes (Tåning, 1952; Lindsey, 1988; Wimberger, 1992; Johnston, 2006; Sfakianakis *et al.*, 2011). Less is known about how early influences can change sex differentiation, later growth rate, life-history traits and behaviour of organisms. These issues did not receive much attention before the theory on life-history evolution and flexibility was developed in the latter part of the 20th century (Roff, 1992; Stearns, 1992). Most empirical knowledge on these effects is therefore recent (Danchin & Wagner, 2010; Salvanes *et al.*, 2013).

Phenotypes emerge from complex interactions between genes and environment (Gilbert & Epel, 2008), and development effects are caused by conditions that organisms experience during embryogenesis (Martin *et al.*, 2013). The earlier the environmental disturbance, the stronger its long-term effects can be (Lindström, 1999). Developmental effects are often overlooked when studying life-history variation, and

<sup>\*</sup>Author to whom correspondence should be addressed. Tel.: +4740480674; email: bror.jonsson@nina.no

phenotypic variation in characteristics, such as growth rate, age at maturity and reproductive investments, can easily be misinterpreted for genetic differentiation. Thus, possible effects of early influences should be considered when studying ecological variation along environmental gradients or between contrasting environments.

A growing body of literature suggests that phenotypic plasticity caused by early experience play a critical role in brain development and flexibility influencing cognition, behaviour, social skills, stress responsiveness and personality development (Malekpour, 2007; Gudsnuk & Champagne, 2011). Also, however, life-history traits, growth and age at developmental shifts are phenotypically plastic and can be affected by early stimuli (Schlichting & Pigliucci, 1998; Jonsson & Jonsson, 2011). For example, poor environmental conditions early in life can give low growth, small adult size and small energy reserves (Taborsky, 2006). Together, such changes reduce the competitive ability of organisms with subsequent negative effects on reproductive success and fitness (Arendt, 2000; Lummaa & Clutton-Brock, 2002). Hence, developmental effects can have paramount importance for understanding life-history and behavioural variation in animal populations. In this review, examples from teleosts and salmonids in particular will be presented. These will illustrate how early environments can influence growth, sex differentiation, life histories and behavioural performances.

The mechanisms behind developmental variation are more abstruse. It is known that environmentally induced change in metabolic rate and growth rate with associated life-history effects can be influenced by thermal and nutritional conditions (Elliott, 1994). Furthermore, the regulation can involve modifications of the epigenotype (Gilbert, 2005; Cabej, 2012) initiating neural, hormonal and enzymatic effects (Gabillard et al., 2005; Rungruangsak-Torrisen et al., 2006; Crews, 2010; Nelson & Monteggia, 2011). It is also known that there are limits for phenotypically plastic variations and that these limits can be inherited (Scheiner & Holt, 2012; Mesoudi et al., 2013). Physiologically, gene expressions can be altered through a neuroendocrine pathway: the nervous system monitors the environment and transfers signals to the endocrine system. The endocrine hormones can then alter gene expression. Alternatively, gene expressions can be changed as an epigenetic effect not altering the underlying DNA sequence. This may be caused by a change in the environmentally induced methylation pattern of the genome, which can alter its transcriptional capabilities. Although epigenetic [epi = over or above (the genetics)] variations are independent of the nucleotide sequence, they can still be inherited over generations (Bossdorf et al., 2008). Other epigenetic mechanisms may be involved, that are mentioned later in this review. There are many uncertainties concerning epigenetic mechanisms influencing phenotypic flexibility, and information on fishes is chiefly restricted to studies of model species such as zebrafish Danio rerio (Hamilton 1822), and medaka Oryzias latipes (Temminck & Schlegel 1846) (Iida et al., 2006; Mirbahai et al., 2011).

This review focuses on how early experiences can affect development and later performance in fishes, *i.e.* how they accomplish given tasks against known standards. The review is mainly limited to influences prior to external feeding, but in a few cases it also includes social influences during the first period of external feeding. Later effects include phenotypic effects visible during later juvenile and adult stages. Characteristics included are juvenile growth and adult body size, sex determination, life-history characteristics such as age at maturity, egg size and fecundity, partial migration, survival

and lifespan. Behavioural aspects covered are social behaviour towards conspecifics, risk taking, anti-predator behaviour, cognition and mate choice.

The presentation commences by limiting the field reviewed. Then, mechanisms by which early influences can be conveyed are presented. The thermal environment is most important in the review of growth, sex determination and life-history effects. Effects of social environments have received most attention in studies of behavioural aspects. Salmonids dominate the examples illustrating life-history effects of early influences, whereas model species are more important in the behavioural aspects covered. At the end of the review, several issues and questions for future studies are suggested.

The review does not penetrate deeply into the molecular or physiological basis for such effects, but mentions theories on how the environment may influence functional phenotypes. The review does not cover anomalies caused by overexposure to temperature, heavy metals or other toxic substances. Environments' effects upon genes are central in oncological research and can influence the initiation and development of cancers (Chik *et al.*, 2011; Dawson & Kouzarides, 2012), but these effects are also outside the scope of this paper.

# EARLY EXPERIENCES WITH EFFECTS ON LATER PERFORMANCE AND LIFE HISTORY

#### WHAT IT IS

Eggs, embryos, larvae and young juveniles can encounter environments with significant effects on their subsequent morphology, physiology, behaviour and life history. For instance, the thermal environment does not only influence the development of specific organs, muscles or other body parts of the embryo, but can also influence ecological traits such as individual growth rate, age and size at ontogenetic niche shifts, age at maturity, fecundity, egg size and lifespan (Table I). All these characteristics can affect subsequent reproductive success and fitness (Harrison *et al.*, 2010).

Developmental effects can be caused by favourable conditions during embryogenesis, which exert positive long-term influences in adult life ('silver spoon effects'). Favourable temperature conditions during the egg development can increase developmental rates and give organisms head starts relative to those developed under less favourable conditions. Advanced developmental rate can reduce age and size at ontogenetic niche shifts with possible consequences for survival and reproductive success (Jonsson, *et al.*, 1990*a*, *b*, 2005; Madsen & Shine, 2000). Such effects are observed in fishes (Koumoundouros *et al.*, 2009), but more studied in reptiles (Warner *et al.*, 2010) and mammals (Descamps *et al.*, 2008).

Behavioural patterns can be based on early experiences and be permanently retained. For instance, male first-generation sea-ranched Atlantic salmon *Salmo salar* L. 1758 are bolder and exhibit prolonged, intraspecific combats compared with wild conspecifics from the same population (Fleming *et al.*, 1996, 1997). Wild *S. salar* grow up in natural rivers where they meet larger and stronger fish, and it can pay to be careful and hide or exhibit subordinate postures and colouration when meeting superior competitors (Hollis *et al.*, 2004; Stuart-Fox & Moussalli, 2008). In contest competition, *S. salar* in rivers perform rapid attacks and retreats and thereby avoid being injured by stronger fish. Sea-ranched *S. salar*, on the other hand, usually grow up in hatchery

TABLE I. Early experiences and types of later performances covered in this review

Early experience	Effect on later performance
Temperature during embryogenesis	Juvenile growth rate
Temperature during egg development	Sex determination
	Adult size
Egg size	Age at maturity
Paternal care	Egg size
Maternal condition and stress	Fecundity
Acidity in water	Survival and lifespan
Early density	Cognition and learning
Predator stress	Boldness and risk taking
	Partial migration

tanks where all competitors are of similar age and size, and there is little risk of serious injuries when fighting over resources. When fed, they rush to the feed and they appear to develop behavioural patterns during their first couple of months linking activity to aggressiveness and exploration tendency (Andriaenssens & Johnsson, 2013). In hatchery-produced salmonids, this behavioural syndrome appears to hold years after being released in nature (Fleming *et al.*, 1997; Réale *et al.*, 2010). Thus, liberated hatchery-produced salmonids are more often injured than wild conspecifics, which exhibit more cautious behaviours (Jonsson, *et al.*, 1990a; Biro & Stamps, 2008), as illustrated in experiments performed in spawning arenas with wild, farmed and sea-ranched *S. salar* (Fleming *et al.*, 1996; 1997).

Similarly, in three-spined sticklebacks *Gasterosteus aculeatus* L. 1758 social behaviours are partly experience based. Aggression from parents on newly hatched offspring can prepare their young ones for proper anti-predation behaviour. Males guard and tend the eggs, but only in habitats with predators, they chase away their offspring a few days after hatching. By this pursuit, they prime adequate anti-predator responses in the developing juveniles (Huntingford *et al.*, 1994). Thus, although genetic differences play an important role in expressions of personality traits (Sundström *et al.*, 2004), parental effects and gene–environment interactions further shape an individual's behaviour (Carere *et al.*, 2005; Stamps & Groothuis, 2010).

The mechanism inducing different social behaviours, such as those between wild and liberated hatchery fishes, is unknown. It was found, however, that cultured Pacific salmonids *Oncorhynchus* spp. have smaller brains than similar-sized wild conspecifics (Kihslinger & Nevitt, 2006; Kihslinger *et al.*, 2006). Furthermore, sensory organs, such as the lateral line system and eyes, can be modified during hatchery rearing and influence behavioural performances (Marchetti & Nevitt, 2003; Anras & Lagardere, 2004). Also, the brain—gene expression profiles in *S. salar* can be affected by the juvenile rearing environment with effects on which behavioural tactics adult fish use (Aubin-Horth *et al.*, 2005). Experiments have demonstrated that a year after hatching, juvenile *S. salar* reared in an impoverished tank environment had lower neural plasticity in the telencephalon and down-regulated neurogenic differentiation in one mRNA expression (transcription factor encoded by the *neuro1* gene) compared to similar juveniles with early experience from an enriched tank environment (Salvanes *et al.*, 2013). The exposure to the enriched environment produced fish with superior learning abilities as

exhibited by enhanced abilities to correctly locate a maze exit during 7 days of testing. Thus, there may be a link between neural plasticity gene expression markers and the spatial behaviour of fishes.

A difference in early social environment can also be the reason for differences in gene expression in the cooperatively breeding cichlid *Neolamprologus pulcher* (Trewavas & Poll 1952). Taborsky *et al.* (2013) compared the expression of four genes of the hypothalamic–pituitary–interrenal (HPI) axis in adults, which had been exposed to two early life social treatments 1.5 years prior to brain sampling. They found that early social experiences altered the programming of the stress axis, and suggested that the adult social behaviour was persistently modified by early life social experience. Behavioural effects of the early rearing environment for *S. salar* and brown trout *Salmo trutta* L. 1758 are currently tested in management-related experiments with the purpose of producing fishes with more natural behaviours (Näslund *et al.*, 2012; Hyvarinen & Rodewald, 2013).

There can also be trans-generational influence caused by environmental conditions encountered by parents resulting in heritable changes in the offspring as in the case of sheepshead minnow *Cyprinodon variegatus* Lacépède 1803. Here, elevated temperature experienced by either parent prior to fertilization modified offspring thermal reaction norms for growth (Salinas & Munch, 2012). It is not known how widespread this epigenetic effect is among fish species, but epigenetic effects are potentially important for offspring development. They may compensate for a poor start under some conditions, and such effects will probably be more keenly studied in the future. At the very least, epigenetic responses may be relevant in relation to climate change effects on organisms (Jonsson *et al.*, 2005; Finstad & Jonsson, 2012).

#### WHAT IT IS NOT

Early experience effects differ from 'seasonal compensation effects' that can occur when there is a reduction in population size in one season that results in a change in per capita rates in the following season. It also differs from compensatory effects where fishes, after periods of food shortage or unfavourable thermal conditions, can exhibit higher than normal growth rates restoring energy reserves (Arendt, 1997; Ali et al., 2003; Gurney et al., 2003). After periods of hunger, individuals can compensate for lost feeding opportunities in a subsequent hyperactive period. For instance, rainbow trout Oncorhynchus mykiss (Walbaum 1792) starved for 3 weeks grew twice as fast as unstarved conspecifics when fed during the subsequent 3 weeks (Dobson & Holmes, 1984). Similarly, Johnsson & Bohlin (2005) showed compensatory growth in S. trutta during 1 month after a period of starvation. Fish kept on the restricted diet restored lost body mass and condition within a month. Compensatory growth is probably a mechanism by which fishes are able to pass energy thresholds associated with developmental switches, such as maturation or migration, and not a compensation for lost growth per se (Álvarez & Nicieza, 2005). Size can be vital in situations where the fish is predator or prey and hence influences the outcome of trophic interactions.

Developmental effects are not 'carry-over effects' which occur when processes in one season influence the success of an individual in the following season. For instance, fishes can increase their reproductive effort when well fed prior to spawning or skip spawning if food is scarce (Bunnell *et al.*, 2007; Kennedy *et al.*, 2008). In *S. trutta* and *S. salar*, male gonad size was found to decrease with migratory costs associated

with spawning (Jonsson & Jonsson, 2003, 2006). Bunnell *et al.* (2007) reported that in white crappie *Pomoxis annularis* Rafinesque 1818 females produced larger ovaries and eggs when better fed prior to maturation. Carry-over effects can be important for environmental adaptability, but are not reviewed here.

#### **MECHANISMS**

Phenotypically plastic reactions may be conveyed by enzymatic influence caused by abiotic or biotic characteristics of the environment. Changes can also be caused by learning, parental influence and through epigenetic effects influencing the physiology of the organism. Epigenetics refer to functionally relevant modifications of the genome that do not involve a change in the nucleotide (DNA) sequence (Holliday & Pugh, 1975; Riggs, 1975; Jones & Taylor, 1980), and may also be provided by the mother as a parental effect, be inherited or based on own experience (Singh *et al.*, 2012). These are all mechanisms with effects on phenotypically plastic reactions. The causes presented in this review are experiences encountered either *via* their parents or during egg incubation or early life, whereas the effects produced last through juvenile life or longer.

#### LEARNING AND MEMORY

Fishes learn from experience and can use the knowledge gained in decision-making. One well-known example of early learning is the salmonid memory of their natal area, which they use to find their way to the spawning grounds in a river they left years earlier for feeding in the ocean (Hasler & Scholz, 1983), but fishes do not only learn the location of their natal area, they also know how to navigate back to their home stream and natal area from distant feeding areas (Hansen et al., 1993), whether this is with help of a cognitive map, magnetic orientation or olfactory cues (Quinn, 1980; Dittman & Quinn, 1996; Putman et al., 2014). During navigation, they may use associations between different stimuli, such as the location of specific resources in relation to the position of landmarks (Warburton, 2003), and they can plan escape routes when threatened by a predator (Ebbeson & Braithwaite, 2012). Also, fishes can form mental maps in which they can encode both vertical and horizontal components (Holbrook & Burt de Perera, 2011), and they can integrate temporal and spatial information and use this to predict when and where specific events will occur (Reebs, 1996). By combining information, they can comprehend ways to respond or behave in complex situations encountered later in life.

There appear to be critical periods when the brain is most plastic and receptive to environmental influences (Knudson, 2004). The critical periods of development occur at key transitions from one life stage to the next, such as early postnatal development and puberty with heightened neural development (McCormick & Green, 2013; Stratman & Taborsky, 2014). Early life is important for the development of sensory processes and cognition where the fishes have to adapt to a range of foraging opportunities, to cope with varying predation threats and social interactions (Ebbeson & Braithwaite, 2012).

#### PARENTAL EFFECTS

Parental effects have traditionally been called maternal effects because effects transmitted *via* the mother were the first to be noticed and because paternal effects, other than

genetic effects transmitted *via* the father, were thought to be minimal or non-existent (Roach & Wulff, 1987; Rossiter, 1996). Recent studies, however, suggest that paternal effects may be more important than previously thought (*e.g.* insects: Giesel, 1986, 1988; Fox *et al.*, 1995; mammals: Clutton-Brock, 1991). Fathers can influence their young ones in ways such as through fertilization success and parental care, and thereby, they indirectly affect offspring competitive ability, growth and associated life-history traits.

A parental effect is any parental influence on offspring phenotype that cannot be attributed to offspring genotype, and to the action of non-parental components of the offspring's environment (Lacey, 1998). Parental effects occur when the phenotype of an individual is determined not only by its own genotype and environmental conditions experienced, but also by the parental phenotype and environment (Wright, 1969). A parental effect can be spawning at a more appropriate time or place, or a physiological effect such as the provisioning of a larger amount or higher quality yolk to the embryo, improving larval growth and survival (Stratman & Taborsky, 2014). It may also have longer-term effects such as influencing age at maturity or lifespan of the offspring (Heath & Blouw, 1998). It can be an epigenetic effect where the function, but not the base sequence of the genome, is changed. Studies have demonstrated that parental effects can have important ecological and evolutionary consequences (Reznick, 1991; Räsänen & Kruuk, 2007; Stratman & Taborsky, 2014).

In most organisms, progeny phenotypes tend to be influenced more by the genotype or environment of their mother than by the genotype and environment of their father (Heath *et al.*, 1999). Studies have demonstrated that parental effects can have important ecological and evolutionary consequences (Reznick, 1991; Räsänen & Kruuk, 2007). Variation in offspring size is one of the best understood mechanisms by which variation among females influences the phenotypes of progeny. In fishes, maternal (relative to paternal) effects are best documented.

In many fish species, juvenile and adult ecologies differ because the individuals grow through life, and thereby, their environmental requirements alter, thus changing their competitive abilities and predator susceptibility. Juveniles and adults of the same species may have more different ecological requirements than adults of different species (Werner, 1977). Therefore, fishes can undergo ontogenetic shifts in feeding niche, and juveniles can occupy a different niche from adults in a common habitat or juveniles and adults can be spatially separated (Werner & Gilliam, 1984; Jonsson & Jonsson, 1993). It is expected that organisms occupy the habitat that minimizes mortality rate over growth rate, rates which vary with time and size of the fish. For adults, precise forecasting of offspring conditions from the ambient environment can be difficult or even impossible if the two live in different habitats. Hence, females may programme their offspring to live in a similar environment they experienced when young, which quite reliably can predict their offspring's future environment. This situation can give rise to 'life stage-specific maternal effects', where the environmental conditions experienced by females during a certain life stage (e.g. the early juvenile stage) induce a maternal effect that affects the same life stage in the offspring generation (Kotrschal et al., 2012).

An example of how maternal experience can influence offspring *via* non-genetic mechanisms is from a study of *G. aculeatus* (Giesing *et al.*, 2011). There, predator-exposed females produced larger eggs with higher cortisol content, and their juvenile offspring swam in tighter shoals (an anti-predator defence) than offspring of

non-exposed females. Thus, mothers exposed to an ecologically relevant stressor during egg formation can transfer a long-lasting effect to their progeny, which may be mediated by exposure to maternally derived cortisol. The results support the hypothesis that female *G. aculeatus* can influence development, growth and behaviour of their offspring *via* eggs to match their future environments.

#### EPIGENETIC EFFECTS

Phenotypic variation can result from an environmental effect on the genotype as an epigenetic change (Roberts & Gavery, 2012). Epigenetic effects can be involved in the differentiation of cell lines and trigger altered gene expression (Jablonka & Raz, 2009), and it can be induced by hybridization or environmental stress and contribute to adaptations to novel environments. Often, epigenetic effects are caused by conditions encountered by the embryo or larva, but can also be trans-generational effects caused by conditions experienced by the parents (de facto parental effect). Experiments with rats as a model organism have demonstrated permanent growth retardation in offspring nursed by mothers fed a low-protein diet (Desai & Hales, 1997). Also, the programming of liver metabolism can be changed as reflected by permanent changes in activities of key hepatic enzymes of glycolysis and gluconeogenesis, which bias the liver towards a 'starved' setting. Desai & Hales (1997) also reported an increase in the life span of offspring exposed to maternal protein restriction during the lactation period, and a decrease in life span when exposed to maternal protein restriction during gestation. The latter study revealed that hepatic metabolism and lifespan can be programmed by events during early life without involving changes in the nucleotide sequence.

The molecular basis of epigenetic effects involves modifications of the activation of certain genes, and proteins associated with DNA may be activated or silenced such as in genomic imprinting, where one of the two alleles of a gene is silenced and expressed in a parent-of-origin-specific manner (Jablonka, 2001). One of the mechanisms is DNA methylation, usually associated with inhibition of genetic transcription.

Changes in DNA methylation, unlike DNA mutations, can be influenced by the environment; they are stable at the time scale of an individual, and present different levels of heritability. These characteristics make DNA methylation a potentially important molecular process to respond to environmental change (Angers *et al.*, 2010). Other epigenetic effects on gene expression include histone- and RNA-based modifications such as microRNA, small interfering RNA, and spatial location of DNA and the organization of the chromatin matrix in the nuclear space, as well as three-dimensional templating mechanisms and self-sustaining loops (Matouk & Marsden, 2008; Richards *et al.*, 2010; Gräff *et al.*, 2011; Khrameeva *et al.*, 2012). Among these, DNA methylation is best studied.

DNA methylation affects many developmental processes responsible for phenotypic variation. The methylation occurs in certain regions of the genome, and refers typically to the addition of a methyl group to position 5 of cytosine bases when cytosine occurs adjacent to a guanine nucleotide (McGowan & Martin, 1997) but it is becoming increasingly evident that there may also be significant methylation in non-cytosine-guanin contexts, at least in pluripotent cell types and oocytes (Ramsahoye *et al.*, 2000; Tomizawa *et al.*, 2011). Addition of methyl groups is catalyzed by enzymes called DNA methyltransferases. DNA methylation stably alters the expression of genes in cells as cells differentiate from embryonic stem cells to specific tissues.

High levels of DNA methylation are associated with silencing of gene expression, and reduced methylation is linked to active gene transcription (Bird, 2002). Temperature can influence the intensity of the DNA methylation, and it occurs abundantly in polar marine species (Kakutani, 2002). The relative amount and location of DNA methylations vary across taxa. It is plausible that these marks can have different functions and different mechanisms of action across organisms and evolutionary time (Roberts & Gavery, 2012). Although implications of DNA methylation are well known, the fundamental understanding of its functioning in fishes is lacking (Simonet *et al.*, 2013).

Histone (proteins that package and order the DNA into structural units called nucleosomes) modifications also affect gene expression. Histone acetylation is a parallel to methylation, but whereas methylation is generally associated with inhibition, acetylation is associated with activation. Histone acetylation is important in regulating the structure and function of chromatin, and hence, the eukaryotic genome. Zhang *et al.* (2013) reported that histone deacetylation was involved in sex differentiation in ricefield eels *Monopterus albus* (Zuiew 1793). They suggested that histone modification in addition to DNA methylation may be widely used to drive natural sex change in teleosts and gonadal differentiation in vertebrates.

Most epigenetic changes occur within the course of one individual organism's lifetime, but they can be inherited trans-generationally (Chandler, 2007). They are the emergent properties of the genome and the response to its environment, both of which are heritable and mutable (Kutschera & Niklas, 2004).

#### EXAMPLES OF TRAITS AFFECTED BY EARLY EXPERIENCES

#### GROWTH AND SIZE

Most studies on effects of early influences on later growth have been performed with endotherms in relation to long-term consequences of early life stress and the resulting glucocorticoid exposure, maternal interactions, and foetal development (de Boo & Harding, 2006; Champagne, 2008). Strangely enough, fewer studies are on ectotherms, although their growth depends on environmental temperature (Schlichting & Pigliucci, 1998; Wootton, 1998) (Table II). In fishes, maximum individual growth rate increases with temperature to an optimum value, and then it decreases rapidly at higher temperatures (Elliott, 1994). The decrease at high temperature is probably because dissolved oxygen in the water decreases with increasing temperature whereas the fish's need for oxygen increases (Brett, 1952; Pörtner, 2010). This function allows ectotherms to maintain thermal homeostasis within a wide range of temperatures (Angilletta *et al.*, 2002). Also, thermal experience from the perinatal period can modulate the normal course of development and permanently alter phenotypes.

Temperature influences the rate of myogenesis (formation of muscular tissue), including composition of the functional structures within the cells, patterns of gene expression, number and size of muscle fibres, fibre diameters and myofibril areas (Johnston, 2006). From studies of *S. salar*, Stickland *et al.* (1988) reported that post-hatching embryos, incubated at higher temperatures, had larger, but fewer muscle fibres, than those incubated at lower temperatures. Large muscle fibres contain more myofibrillar material than smaller fibres. Campos *et al.* (2013) reported similar findings from studies of Senegalese sole *Solea senegalensis* Kaup 1858, and Steinbacher *et al.* (2011)

TABLE II.	Early	influence	with	delayed	effects	on	juvenile	growth.	Experimental	species and	
study cited are given											

Influence	Experimental species	References
Embryonic temperature	Rutilus meidingeri Salmo salar Danio rerio	Steinbacher <i>et al.</i> (2011) Finstad & Jonsson (2012) Scott & Johnston (2012)
Early predator stress Parental care Maternal maturation temperature Amount of yolk	Oncorhynchus mykiss Amphiprion melanopus Cyprinodon variegatus Simochromis pleurospilus	Bell <i>et al.</i> (2011) Green & McCormick (2005) Salinas & Munch (2012) Segers <i>et al.</i> (2012)

found that pearlfish *Rutilus meidingeri* (Heckel 1851) incubated at high temperature were larger at hatching than those incubated in colder water. The latter had reduced cell differentiation leading to smaller hatchlings, which secured a larger pool of muscle precursor cells used for enhanced post-hatchling growth, finally resulting in larger adults.

Somatic growth is determined by the surplus energy of the organism, i.e. energy not used for activity, maintenance and specific dynamic action. The cost of maintenance is influenced by the standard metabolic rate (Burton et al., 2011), and Álvarez et al. (2006) reported that the standard metabolic rate of juvenile S. trutta changed in response to temperature during larval development, which parallels the observation that subsequent juvenile growth varies depending on temperature during embryogenesis. This relationship holds for S. salar and some other species (Finstad & Jonsson, 2012). Juvenile S. salar developed from eggs incubated during winter in unheated water (2-4° C) grew less well in water warmer than 17° C than conspecifics with embryogenesis in water heated to  $6-8^{\circ}$  C. This finding explains the observation that cohorts of S. salar incubated during relatively warm winters grew better as juveniles in fresh water and migrated at a younger age to the sea, than those with embryonic development during colder winters. This difference in juvenile growth was independent of the water temperature in the subsequent summer (Jonsson et al., 2005). This early effect on juvenile growth might have been considered in a recent experiment with Atlantic cod Gadus morhua L. 1758. Fish reared from young ones caught in the warmer Clyde Sea on the west coast of Scotland grew faster than those from the colder St Andrews Bay, when the young ones were brought together under similar controlled conditions (Harrald et al., 2010). The authors viewed this as an example of genetic differentiation between populations, but an environmentally induced epigenetic effect would have given a similar result. Thermal effects during egg incubation can last much longer than through the larval period and should be taken into consideration when comparing population-specific adaptations of fishes. This view is supported by the observation from another gadid, haddock *Melanogrammus aeglefinus* (L. 1758); Martell et al. (2005) reported that temperature during embryogenesis influenced subsequent growth. Similarly, Korwin-Kossakowski (2008) reported that growth of young common carp Cyprinus carpio L. 1758 was enhanced by incubating the eggs in relatively warm water, indicating that this may be a common phenomenon in teleosts.

Egg incubation temperature also influences later thermal acclimation. Scott & Johnston (2012) used *D. rerio* as a model species. They observed that temperature during embryogenesis influenced the thermal reaction norm for growth, and that differences

in expression were observed for individual genes involved in energy metabolism, blood vessel formation (angiogenesis), cell stress and muscle contraction. Also, predatory stress can influence later growth. Juvenile *G. aculeatus* exposed to cues of an *O. mykiss* predator grew faster, but were smaller at adulthood than corresponding juveniles prevented from exposure to *O. mykiss* (Bell *et al.*, 2011). The smaller size probably resulted from a deferred cost of early rapid growth with subsequent reduced age and size at maturity (Alm, 1959; Jonsson *et al.*, 2013).

Maternal and paternal effects can influence size and growth of fishes. In tropical clownfish *Amphiprion melanopus* Bleeker 1852, Green & McCormick (2005) observed that larval size at metamorphosis was primarily affected by maternal and paternal effects. Paternity explained 52% of the variance in growth rates, while the combination of paternity, maternity and temperature explained 30%. This strong paternal influence may be due to the extensive role males play in nest-tending coupled with the relatively long embryonic duration of the species.

Trans-generational thermal effects experienced by the parent prior to egg fertilization can also translate into significant changes in the shape of offspring reaction norms for growth (Mousseau & Fox, 1998). Salinas & Munch (2012) reported that female C. variegatus kept at either 24, 29 or 34° C for 30 days during egg maturation produced offspring that grew best at the temperature experienced by their mothers. The change in growth over a single generation (c. 30%) exceeded the single-generation rate of adaptive evolution by an order of magnitude. In this species, mothers adaptively programmed their offspring for maximal growth at the present temperature.

Growth hormone expressions can also influence juvenile growth. Segers *et al.* (2012), using the mouth breeding cichlid *Simochromis pleurospilus* Nelissen 1978 as a model species, reported that young ones originating from small eggs had higher growth hormone receptor expression levels causing juveniles from small eggs to grow faster than those from larger eggs. A similar relationship may explain Berejikian *et al.*'s (2001) finding that offspring growth rate and egg size in Chinook salmon *Oncorhynchus tshawytscha* (Walbaum 1792) were negatively correlated.

Presence of specific metabolic enzymes, such as different trypsin isozymes, can influence growth. Trypsin is a key digestive protease, which is sensitive to environmental change and influences feed utilization and growth during the life cycle (Rungruangsak-Torrissen & Male, 2000). The trypsin isozyme variants influence maintenance ration and the capacity for protein synthesis in white muscle with effects on growth, size and other life-history variables. The genetic mechanism for flexible trypsin production is unknown, but Jump & Clark (1999) maintained that environmental factors may interact with the genome and allow cells to adjust to an on and off switch gene expression, possibly influenced by DNA methylation. Sainz *et al.* (2005), on the other hand, concluded from a study of isotrypsin patterns that the regulation was quantitative not qualitative, and that changes in trypsin activity and in trypsin mRNA by internal and external stimuli must be related to changes in the rate of synthesis of the established isoenzymes. In any case, trypsin regulation is one possible mechanism by which growth is influenced by temperatures experienced by the embryos.

From endotherms, it has been observed that maternal hormonal responses can alter the offspring phenotype (Kaiser & Sachser, 2009). For instance, maternal androgens deposited in avian eggs provide a flexible mechanism of non-genetic inheritance by which the mother can adjust the development of some over the others

(Groothius *et al.*, 2005). In fishes, the physiological condition of mothers during gametogenesis can influence offspring quality of damselfish *Pomacentrus amboinensis* Bleeker 1868 (McCormick, 1998). The reason may be that competitors and predators can elevate maternal cortisol levels and consequently influence larval morphology through a stress-related response. Thus, behavioural interaction regimes can determine larval quality and potentially govern a female's contribution to the next generation.

#### SEX RATIO

Environmentally influenced sex determination has stimulated much research. In most fish species, sex is genetically determined, but in some, environmentally influenced sex differentiation has been documented. Water temperature is the main environmental determinant of sex, and also water acidity (Heiligenberg, 1965; Rubin, 1985; Francis & Barlow, 1993), fish density and social interactions, growth rate and contaminants can affect sex ratios (Baroiller & D'Cotta, 2001; Baroiller *et al.*, 2009*a*; Sandra & Norma, 2010; Siegfried, 2010) (Table III). Thermal influences are best studied.

# Thermal influence

Thermal sex determination occurs in several species. In some cases, the sex is changed if eggs or larvae are exposed to extremely high temperatures, but not under normal conditions, such as in sockeye salmon *Oncorhynchus nerka* (Walbaum 1972), O. mykiss, O. latipes, Nile tilapia Oreochromis niloticus (L. 1758) (Tessema et al., 2006) and D. rerio. In other cases, sex is influenced by temperatures individuals typically encounter in nature. This is better known from reptiles where temperature influences sex determination by modulating the activity of steroidogenic enzymes affecting the hormonal environment of the embryo inducing masculinization or feminization (Janzen, 1994), but it also occurs among teleosts. Ospina-Álvarez & Piferrer (2008) reported temperature-influenced sex differentiation in 33 cichlids and a few other species (Table III). In flatfishes of the genus *Paralichthys*, larvae of genetic females may exhibit sex reversal when exposed to warm water (Montalvo et al., 2012a, b). Also, O. niloticus may exhibit sex reversal in natural lakes. The temperature sensitivity is under genetic control, and surplus males in temperature-treated groups can be selected as a quantitative trait (Rougeot et al., 2008; Baroiller et al., 2009b; Lozano et al., 2013). Recently, Wedekin et al. (2013) reported that the frequency of male European grayling Thymallus thymallus (L. 1758) increased with increasing temperature in Lake Thun, Switzerland, and the authors speculate about thermally influenced sex determination in this species.

In cases of thermally influenced sex differentiation, the sex is determined during a sensitive period of early development (Valenzuela *et al.*, 2003). Typically, species without consistent genetic differentiation produce relatively more males as the temperature increases (Ospina-Álvarez & Piferrer, 2008; Hasyashi *et al.*, 2010), but in the channel catfish *Ictalurus punctatus* (Rafinesque 1818) (Patino *et al.*, 1996), Korean rockfish *Sebastes schlegeli* Hilgendorf 1880 (Omoto *et al.*, 2010), European seabass *Dicentrarchus labrax* (L. 1758) (Baroiller & D'Cotta, 2001) and possibly in dwarf perch *Micrometus minimus* (Gibbons 1854) high temperatures induce relatively more females. In the last species, Schultz (2008) observed that broods developing at low temperatures early in the season produced relatively more males than those developed in warmer water later in the season. In the olive flounder *Paralichthys olivaceus* 

TABLE III. Environmental variables affecting sex determination in fishes

Variables	Species	References
Water acidity	Pelvicachromis pulcher Pelvicachromis taeniatus	Rubin (1985)
	Pelvicachromis taentatus Pelvicachromis subcellatus	
	Apistogrammus borelli	
	Apistogrammus cacatuoides	
	Apistogrammus diplotaenia	Römer & Beisenherz (1996)
Temperature	Apistogrammus spp.	Römer & Beisenherz (1996); Ospina-Álvarez & Piferrer (2008)
	Menidia menidia	Ospina-Álvarez & Piferrer (2008)
	Menidia penninsulae	
	Odontesthes bonariensis	
	Odontesthes argentinensis	
	Haplosterum littorale	
	Poeciliopsis lucida	
	Limia melanogaster	
	Oncorhynchus nerka	Craig <i>et al.</i> (1996); Azuma <i>et al.</i> (2004)
	Oncorhynchus mykiss	Magerhans et al. (2009)
	Oryzias latipes	Selim et al. (2009)
	Oreochromis niloticus	Tessema et al. (2006)
	Danio rerio	Abozaid <i>et al.</i> (2012); Villamizar <i>et al.</i> (2012)
	Ictalurus punctatus	Patino et al. (1996)
	Sebastes schlegeli	Omoto et al. (2010)
	Dicentrarchus labrax	Baroiller & D'Cotta (2001)
Growth	Danio rerio	Lawrence et al. (2008)
	Rutilus rutilus	Paull et al. (2009)
Density	Macropodus opercularis	Baroiller & D'Cotta (2001)
Growth, density or social influence	Anguilla spp.	Krueger & Oliveira (1999); Davey & Jellyman (2005)
	Paralychthys lethostegma	Luckenbach et al. (2003)
	Ichthyomyzon gagei	Beamish (1993)

(Temminck & Schlegel 1846), both high and low temperatures induce monosex male populations while intermediate temperatures yield a 1:1 sex ratio (Baroiller & D'Cotta, 2001).

The effect of temperature on sex determination can depend on whether temperature is stable or varies between day and night. Villamizar *et al.* (2012) found that temperature varying between 24 and 28° C during day and night gave more females in *D. rerio* than at either temperature when stable.

# Physiological regulation

In gonochoristic fishes, steroid hormones, oestrogens in females and 11-oxygenated androgens in males regulate key physiological steps of gonadal sex differentiation

(Baroiller & D'Cotta, 2001). The temperature-sensitive cytochrome P450-aromatase enzyme catalyzes the oestrogen biosynthesis and feminization by conversion of androgens to oestrogens as, for example, demonstrated for southern flounder Paralichthys lethostigma Jordan Gilbert 1884 (Montalvo et al., 2012a, b). Furthermore, in O. niloticus and P. olivaceus, the aromatase gene expression is down-regulated by masculinizing temperature treatments. Masculinization at high water temperature has been ascribed to stress and elevated cortisol concentration during gonadal sex differentiation, as in larval P. olivaceus (Yamaguchi et al., 2010). Cortisol may promote the level of 11-ketotesterone, the main androgen in fishes, and thereby induce the masculinization as demonstrated for Argentinian silverside Odontesthes bonariensis (Valenciennes 1835) (Fernandino et al., 2012). In Atlantic silverside Menidia menidia (L. 1766), Blazquez & Somoza (2010) suggested that an aromatase enzyme encoded by the cyo19a1b gene may be involved in this process. A form of this enzyme occurs in high concentrations in the brain of teleosts, and may be involved in the conversion of androgens into oestrogens and thereby in feminizing fishes at low temperature. In experiments with D. labrax, it was observed that at high temperatures juvenile males have double the DNA methylation levels of females in the promotor region of the gonadal aromatase (Navarro-Martin et al., 2011). Exposure to high temperatures increased the promotor methylation levels of females inhibiting development of ovaries, and DNA methylation at high temperature may be an essential component of the mechanism of temperature-dependent sex determination of the species. As mentioned earlier, histone deacetylation appears to be involved in sex differentiation of M. albus (Zhang et al., 2013).

#### Ecological significance

The adaptive significance of environmentally induced sex differentiation is not well understood. One possibility is that it is a remnant trait still present and retained because the cost is small relative to the more common genetic sex determination. It is assumed, however, that environmental sex reversal overpowering genetic sex determination probably has fitness costs. Senior *et al.* (2012) meta-analytically combined the results of 78 studies of chemically induced sex reversal, and found that the fitness of the fish was reduced at the time of exposure. Exposed individuals exhibited a smaller size and a smaller gonado-somatic index than unexposed individuals. This result held independent of growth rate of the fish.

Charnov & Bull (1977) predicted that selection would favour environmentally induced sex determination over chromosome-based sex determination when the fitness of the sex varies among environments. This model appears to hold for *M. medinia* (Conover, 1984). There, offspring produced early in the breeding season maximize relative fitness if they become female, and offspring produced late in the breeding season maximize relative fitness by becoming male, and populations at different latitudes compensate for differences in thermal environment and seasonality by adjusting the response of sex ratio to temperature (Conover & Heins, 1987). Furthermore, Conover *et al.* (1992) reported evidence suggesting that environmental sex determination can evolve in response to selection. Also for reptiles, there is some evidence that this mechanism can be adaptive (Warner & Shine, 2008), but for most species, it is an open question if and to what degree environmentally influenced sex differentiation is adaptive or not.

TABLE IV. Early influences with delayed effect on life-history variables. Experimental species and study cited are given

Influence	Characteristic influenced	Species	References
Parental density	Egg size	Heterandria formosa	Leips et al. (2009)
Early growth	Egg size	Salmo salar	Jonsson et al. (1996)
		Salvelinus leucomaenis	Morita et al. (1999)
		Oncorhynchus masou	Tamate & Maekawa (2000)
		Coregonus albula	Gregersen et al. (2011)
Early growth	Lifespan	Gasterosteus aculeatus	Inness & Metcalfe (2008); Lee <i>et al.</i> (2013 <i>b</i> )
Early growth	Partial migration	Salmo trutta	Olsson et al. (2006)
Early spawning	Partial migration	Morone americanus	Kerr & Secor (2010)
Maternal feeding	Egg size	Simochromis pleurospilus	Taborsky (2006)
Maternal predator experience	Egg size	Eretmodus cyanostictus	Segers & Taborsky (2012)
•		Gasterosteus aculeatus	Giesing et al. (2011)

#### LIFE-HISTORY TRAITS

This section summarizes effects of early experiences on life-history traits (Table IV). One of the most well-known examples is that of increased early growth rate that leads to earlier age-at-life-stage transitions such as age when a fish exhibits dietary change or shifts to a better feeding habitat and seaward migration in anadromous species (Jonsson & Jonsson, 1993; Jonsson *et al.*, 1999). Another such transition is during maturation of the fish.

#### Age at maturity

Early maturation is related to juvenile growth, size and fat storage, where both fast initial growth and high lipid density reduce age at maturity. This was demonstrated for a number of freshwater species by Alm (1959) and Day & Rowe (2002) and particularly well documented for salmonid species, *e.g.* Arctic charr *Salvelinus alpinus* (L. 1758) (Jonsson & Hindar, 1982; Forseth *et al.*, 1995), *S. trutta* (Alm, 1959; Jonsson, 1985), *S. salar* (Thorpe, 1986; Berglund, 1992), masu salmon *Oncorhynchus masou* (Brevoort 1856) (Silverstein *et al.*, 1997), *O. tshawytscha* and coho salmon *Oncorhynchus kisutch* (Walbaum 1792) (Vøllestad *et al.*, 2004). In salmonids, winter growth and fat content in the diet as well as genetic constitution also influence age at maturity (Gjerde, 1984; Jonsson *et al.*, 2012, 2013). Thus, factors influencing early growth rate can be reflected in age at maturity.

#### *Egg size and fecundity*

It has been reported that females growing fast at early age produce more, but smaller eggs, than more slow growing conspecifics in *S. salar* (Jonsson *et al.*, 1996), *S. trutta* (Lobon-Cervia *et al.*, 1997), *O. masou* (Tamate & Maekawa, 2000), white-spotted charr *Salvelinus leucomaenis* (Pallas 1814) (Morita *et al.*, 1999) and European vendace

Coregonus albula (L. 1758) (Gregersen et al., 2011), but no such effect was found in O. tshawytscha, O. kisutch and O. mykiss (Quinn et al., 2004, 2011). This discrepancy may indicate that it is not growth per se, but a factor stimulating growth, which caused the observed variation in egg size. As discussed earlier in this review, temperature during embryogenesis enhances juvenile growth and is regularly used in hatcheries to advance developmental rates (Finstad & Jonsson, 2012).

Relationships between egg size, fecundity and growth history are also revealed by studies of latitudinal clines. In S. trutta, O. kisutch and O. masou, egg size is typically larger and egg number lower at more northerly latitudes where early growth is lower (Fleming & Gross, 1990; Jonsson & Jonsson, 1999; Morita et al., 2009). This correlation, however, may not be consequential, but rather, influenced by factors influencing growth rate, such as early temperature. Bownds et al. (2010) tested whether optimal offspring size differs between thermal environments, and if this provided a direct way of assessing the adaptive significance of temperature-driven variation in egg size in D. rerio. The effects of offspring size on performance were highly variable among life-history stages and dependent on the thermal environment; offspring size positively affected performance at some temperatures, but negatively at others. These data were used to generate a simple optimality model; the model predicted that mothers should produce the largest offspring at the lowest temperature, offspring of intermediate size at the highest temperature and the smallest offspring at the intermediate temperature. An experiment showed that the rank order of observed offspring sizes produced by mothers matched the predictions. The mothers adaptively manipulate the size of their offspring in response to thermally driven changes in offspring performance and highlight the utility of optimality approaches for understanding offspring size variation. Also, Iguchi (2012) examined egg size variation. Two populations of ayu *Plecoglossus* altivelis (Temminck & Schlegel 1846) were studied from the Lake Biwa system, Japan, where two streams with different temperatures showed similar maternal body sizes and hatchling sizes, but the eggs were larger in the colder stream. An experiment that used eggs from a single clutch showed that smaller hatchlings were obtained with a lower incubation temperature, indicating that the effect of differences on hatchling size can be cancelled out by variations in incubation temperature.

Lee *et al.* (2013*a*) tested if accelerated or decelerated early growth, caused by a short-term change in temperature, had fitness costs for *G. aculeatus*. Although the fish at adulthood were equal in size, those compensating for low growth by a subsequent accelerated growth rate had reduced reproductive investment (males: reduced sexual ornaments and speed of building nests; females: reduced clutch size, egg size and egg production per year). Thus, *G. aculeatus* with a decelerated growth trajectory showed improved breeding allocation.

Lobon-Cervia *et al.* (1997) reported that the number of eggs of first-time spawners was fixed during their first year of life, whereas the egg size was influenced by second-year growth. Also, according to Morita *et al.* (1999) and Tamate & Maekawa (2000), fast growth during the second year was associated with large eggs in the adult fishes, but the latter authors did not observe any associated effect on egg number. The influence of early growth history on reproductive traits is not only confined to salmonids, but also reported from other species. In a study of *S. pleurospilus*, Taborsky (2006) found that females raised in a poor environment produced larger young ones than females raised without food limitations. This maternal boost was due to a higher investment in eggs and to faster larval growth. As in *S. salar*, mothers appear to prepare

their offspring for rearing conditions similar to those they encountered as juveniles. This may be an adaptation in some species where the juvenile and adult environments are very dissimilar (Jonsson *et al.*, 1996; Taborsky, 2006; Burton *et al.*, 2013). It may also be advantageous for individuals colonizing new habitats with a different thermal regime so that they can modify their reproductive allocation to the novel conditions. Moreover, in a gradually changing climate, this phenotypically plastic mechanism can produce a more rapid change in reproductive investment than can be obtained through natural selection.

The behavioural mechanisms responsible for context-dependent fitness differences were studied in *S. pleurospilus* (Segers & Taborsky, 2011). Larger eggs gave rise to larger young ones with higher burst swimming speed. Food ration greatly influenced long-term growth, while egg size predominantly affected early fish size. Egg size and food ration interactively affected hiding and foraging behaviours of the fish. In the low-food treatment, individuals from small eggs spent less time in shelter and exhibited a higher commitment to foraging than individuals from large eggs. In a natural setting, this should increase predation risk of young ones originating from small eggs, particularly in hostile and nutrient-poor environments. When food was abundant, juveniles behaved similarly, irrespective of egg size. Thus, egg size affected juvenile growth and behaviour differently in different environments. While it is well established that large egg size raises offspring fitness in harsh environments, this study suggests that this advantage arose through risk-averse behaviour linked to offspring size.

Egg size is known to affect growth rate of young *S. alpinus*, and possibly also behaviour, but these effects appear to decrease with time and are minimized by the effects of social factors (Leblanc *et al.*, 2011). Fish hatching from large eggs were more mobile and foraged more than fish developed from smaller eggs. Social environment also affected foraging behaviour, mobility and space use. Group-living *S. alpinus* were more mobile, foraged more and responded faster to food delivery than isolated fish.

On the other hand, fishes do not always produce large eggs in hostile environments (Morrongiello *et al.*, 2012). Southern pygmy perch *Nannoperca australis* Günther 1861 inhabit a diversity of streams along gradients of environmental quality, variability and predictability. Populations inhabiting increasingly harsh streams produced more numerous and smaller eggs. The within-female egg size variability increased as environments became more unpredictable. The authors argued that egg size had only a minor influence on offspring fitness in harsh environments. Instead, maternal fitness was maximized by producing many eggs. The increased variability in size may be an example of bet hedging. Koops *et al.* (2003) found support for the hypothesis that egg size variability increased as the environmental predictability decreased. Einum & Fleming (2004), however, maintained that within-clutch variation, at least in salmon, is more likely a reflection of the direct influences of the rearing environment on a female's ability to allocate resources evenly, and is not an evolutionary adaptation.

Predator stress may also influence the size of the eggs produced. Segers & Taborsky (2012) reported that predator cues perceived by the mouth breeding striped goby cichlid *Eretmodus cyanostictus* (Boulenger 1898) produced heavier eggs, but smaller clutches, than conspecific females without prior predator experience. Thus, also in this species, mothers appear to have the ability to prepare offspring for challenges they may meet after birth by adjusting the amount of yolk in their egg, and thereby maximizing their fitness. Similarly, in *N. pulcher*, the mothers adjust the size of propagules to the selective forces to which the offspring can be exposed. When offspring are cared for by

parents or alloparents, protection may reduce the predation risk for the young ones, which may allow mothers to invest less per single offspring. In *N. pulcher*, brood care helpers protect group offspring and reduce the latters' mortality rate. With more helpers present, females can reduce their investment per egg (Taborsky *et al.*, 2007).

Is there any paternal effect on offspring success? Apparently, it is present in *D. rerio* (Uusi-Heikkila *et al.*, 2012). In the laboratory, females and males from four size groups were allowed to spawn freely. As expected, larger females produced more eggs and larvae with larger yolk sac than smaller females, but male body size further contributed to *D. rerio* reproductive success: offspring sired by large males exhibited higher hatching probability and these offspring hatched earlier and larger than offspring fertilized by small males. The largest males, however, experienced lower mating success and received fewer eggs than males of the smaller size classes. Although little studied, male size may be more important for offspring success than previously expected.

# Partial migration

Partial migration is the divergence of a population into migratory and resident contingents of the same species (Jonsson & Jonsson, 1993). It has been demonstrated that early growth influences whether a fish from such populations becomes migratory or not. For instance, Olsson et al. (2006) showed experimentally with reciprocally transplanted S. trutta between two sections in a river that migratory behaviour developed depending on early density and early individual growth rate. In river sections with high S. trutta densities and low specific growth rates, the fish tended to become migratory. Conspecifics from low density sections where the initial growth rate was high tended to stay resident. This result was confirmed in rearing experiments with variable feeding regimes showing that the migration of S. trutta was environmentally induced (Wysujack et al., 2009). Furthermore, from studies on white perch Morone americana (Gmelin 1789), Kerr & Secor (2010) reported that the migratory fish tended to originate from early spawned larval cohorts characterized by slow growth compared with later spawned cohorts with fast initial growth as revealed by otolith microstructure. Possibly, there are latent effects on partial migration of M. americana related to spawning phenology, its interaction with temperature and resultant larval growth rates. The migratory propensity may be controlled by a threshold switch-point that determines whether a migratory or a resident phenotype is expressed, and growth rate during early life may be one characteristic influencing the choice made. Decisions about migration, however, may be also taken at later occasions depending on the energetic state of the individuals (Forseth et al., 1999).

#### Survival and lifespan

There is a positive correlation between early growth rate and lifespan (Metcalfe & Monaghan, 2003; Inness & Metcalfe, 2008). Faster growth in early life is associated with reduced length of life span. For instance, Lee *et al.* (2013*b*) exposed *G. aculeatus* briefly in cold or warm water to deflect young fish from their normal growth trajectories and showed that increased early growth reduced the life span while growing more slowly prolonged lifespan.

Maternal condition may also influence offspring survival. In the coral reef fish *Acanthochromis polyacanthus* (Bleeker 1855), good condition parents produced progeny with increased survival compared to parents in poorer condition

(Donelson *et al.*, 2008). The offspring from parents in good condition were larger at hatching, had larger yolk reserves and exhibited increased survival. Larger size is expected to provide benefits to offspring through reduced susceptibility to size-selective mortality. The range of offspring characteristics modified by parental condition can thus result in a greater proportion of offspring from good condition parents recruiting to the population.

#### BEHAVIOUR

This section summarizes behavioural effects of early influences (Table V). Some of these effects, such as those of social and predator avoidance behaviour, have growth and life-history consequences which were dealt with in preceding sections.

## Social behaviour towards conspecifics

Phenotypic traits of parents can affect patterns of development in their offspring (Mousseau & Fox, 1998; McAdam et al., 2002). This may be because the progeny develop in an environment chosen by their parents or social learning from the behaviour of their parents (Rossiter, 1996). Rearing in an enriched environment facilitates learning about novel prev items in S. salar (Brown et al., 2003) and G. morhua (Braithwaite & Salvanes, 2005; Salvanes & Braithwaite, 2005), and competitive ability and behavioural variation in O. mykiss (Berejikian et al., 2001; Lee & Berejikian, 2008). Experience of a varying and changeable environment appears to teach the fish flexible behaviours. Also, social learning with effects on the behaviour of the adults can be influenced by density at early rearing. Experiments with guppies *Poecilia reticulata* Peters, 1859 showed that individuals kept at low density during early life had greater shoaling tendency and more effective use of socially learned information. They were quicker to locate food in a maze task and had better learning skill than conspecifics reared at higher density (Chapman et al., 2008). Poecilia reticulata reared at higher densities were less effective because of poorer capacity for watching and learning from shoal mates. Hatchery S. trutta are typically reared at high densities, but in an experiment where the fish were reared at lower, more natural densities, they were twice as likely to survive after release into a natural stream (Brockmark & Johnsson, 2010). Developing in a less crowded juvenile environment had a positive effect on the cognitive ability and overall survival to adulthood.

## Anti-predator behaviour and risk taking

Predator avoidance behaviour can be influenced by early experience (Goodey & Liley, 1986). Experiments have shown that *P. reticulata* chased when young by adult conspecifics required significantly more attacks before they were captured than either *P. reticulata* without early experience of being chased or *P. reticulata* exposed to only visual or chemical cues associated with chasing by adults. Thus, conditioning of predator-avoidance behaviour by early exposure to chasing by potential predators, may be an advantage in physically and biotically heterogeneous environments occupied by *P. reticulata*, and chasing by adult conspecifics may provide part of the learning experience (cf. survival and lifespan).

Prey species may learn the identity of the predator early in life and use this when later approached by the predator (Lönnstedt *et al.*, 2012). Both visual and olfactory

TABLE V. Early influences with delayed effect on later behaviour. Experimental species and study cited are given

Influence	Characteristic influenced	Species	References
Variable feeding	Learning ability	Simochromis pleurospilus	Kotrschal & Taborsky (2010)
Rich rearing environment	Learning ability	Salmo salar	Brown et al. (2003)
		Gadus morhua	Braithwaite & Salvanes (2005); Salvanes & Braithwaite (2005); Strand <i>et al.</i> (2010)
Rich rearing environment	Competitive ability	Oncorhynchus mykiss	Berejikian <i>et al.</i> (2001)
	Behavioural variation	Oncorhynchus mykiss	Lee & Berejikian (2008)
Early habitat complexity	Navigation skill	Salmo salar	Salvanes et al. (2013)
Unpredicatble early environment	Boldness and exploratory behaviour	Poecilia reticulata	Chapman <i>et al.</i> (2010)
Early density	Shoaling tendency	Poecilia reticulata	Chapman <i>et al</i> . (2008)
Early chased and predator experience	Increased predator avoidance	Poecilia reticulata	Goodey & Liley (1986)
*		Pomacentrus wardi	Lönnstedt <i>et al</i> . (2012)
Maternal predator experience	Decreased predator avoidance	Gasterosteus aculeatus	McGhee et al. (2012)
Maternal stress	Decreased feeding efficiency	Salmo salar	Eriksen et al. (2011)
Maternal stress	Schooling	Gasterosteus aculeatus	Giesing <i>et al.</i> (2011)
Adult presence	Increased aggressiveness and submissiveness	Neolamprologus pulcher	Arnold & Taborsky (2010)
Foster mother of another species	Mate preference	Haplochromis pundamilia Haplochromis nyererei	Verzijden & Cate (2007)

cues help the prey recognize the predator. Two to three week old coral reef damselfish *Pomacentrus wardi* Whitley 1927 were exposed to a high or low food ration and conditioned to recognize the olfactory cues (odours) and visual cues from two common benthic predators, the moon wrasse *Thalassoma lunare* (L. 1758) and sand lizard-fish *Synodus dermatogenys* Fowler 1912. Feeding history strongly influenced their

willingness to take risks in the natural environment. Conditioning in the laboratory with visual, olfactory or both cues from predators led fishes in the field to display risk-averse behaviours compared with fishes conditioned with sea water alone.

On the other hand, *G. aculeatus* maternal exposure to predation risk decreases off-spring anti-predator behaviour and survival (McGhee *et al.*, 2012). Gravid females were exposed to a model predator or not, and the offspring's behaviour and survival were compared when they were alone with a live predator, the same type of predator that threatened their mothers. Offspring of predator-exposed mothers were less likely to orient to the predator and were more easily caught than offspring from unexposed mothers. The strength and direction of the maternal effect on offspring survival, however, depended on offspring size. Specifically, the larger the offspring from predator-exposed mothers, the more vulnerable they were to predation compared to offspring from unexposed mothers. These stress-mediated maternal effects can have non-adaptive consequences for offspring when they find themselves alone with a predator.

Inherited inter-individual variation in metabolic rate exists in a wide range of species, and there is a relationship between metabolic rate and risk taking in *D. labrax* (Killen *et al.*, 2012), but boldness and risk taking also have strong environmental components that affect the economy and adequacy of individual reactions to social challenges (Brown *et al.*, 2007). Environment may influence aggressive and submissive behaviours, linked to boldness through an early behavioural syndrome (Andriaenssens & Johnsson, 2013). In experiments with *N. pulcher*, Arnold & Taborsky (2010) showed that juveniles raised with adults exhibited more aggressive and submissive behaviours to each other than those raised with siblings only. The strength of the treatment effects depended on the opponent's social experience, and contests were terminated earlier only when both opponents had been raised with adults.

Unpredictability in early food supply can induce boldness in fishes as demonstrated by Chapman *et al.* (2010) in experiments with *P. reticulata*. Fishes with early experience of unpredictable environments were bolder and more exploratory than those reared in predictable environments. Individuals with early experience of unpredictability spent significantly less time associating in shoals with conspecifics. Also, early density can influence the tendency to shoal.

Effects of early rearing on breeding performance were studied by Fleming et al. (1997). They used S. salar in semi-natural spawning arenas by use of a split experimental design and showed that fish deprived of an early social environment with stronger competitors and predators did not retreat from competition when they met stronger contesters. In their experiments, they matched wild and sea-ranched S. salar of similar genetic background. The sea-ranched fish were tank reared in a hatchery without contact with larger fish. They were liberated at age 1 year when they migrated to sea for feeding in the North Atlantic Ocean together with natural river-produced juveniles. Thus, the sea-ranched fish were deprived of early, juvenile river experience. They did not compete for food among fish of different sizes and strengths. Both the natural and sea-ranched S. salar returned from the ocean 1 year later for spawning in the River Imsa. Although levels of aggressive behaviour were similar, the sea-ranched hatchery males were involved in more prolonged aggressive encounters and incurred greater wounding and mortality than wild males. Furthermore, the hatchery-produced males were less able to monopolize spawnings, and as a result, they only achieved 51% of the reproductive success of similar sized wild males. This reproductive inferiority varied directly with the male density and bias in the sex ratio, reflecting the intensity of male

breeding competition. A lower intensity of female than male competition was probably responsible for the lack of differences in breeding performance between hatchery produced and wild females.

In S. salar, maternal stress can affect the boldness and social dominance of the progeny. Eriksen et al. (2011) simulated maternal stress by giving the females intraperitoneal cortisol implants 1 week prior to fertilization of the eggs. Behavioural tests on the offspring 1.5 years after hatching demonstrated that those from the cortisol-implanted females made more unsuccessful feeding attempts than offspring from control females. There was no difference in social dominance between controls and the progeny of cortisol-treated fish. There were, however, higher proportions of individuals with bite marks in the cortisol group. Offspring of the cortisol-treated females that gained high dominant ranks were more aggressive than control winners. During acute confinement stress, offspring from cortisol-implanted females also showed a reduction in the proportion of time they were moving compared with the controls. Thus, the maternal endocrine state at spawning can affect several aspects of progeny behaviour.

# Cognition

Increased habitat complexity can promote the cognitive ability of fishes. *Gadus morhua* reared in tanks with cobble stones and artificial kelp on the bottom was more adept at learning how to forage on novel prey than those reared in bare tanks (Strand *et al.*, 2010). Also, habitat enrichment improved navigation skills in *S. salar* (Salvanes *et al.*, 2013). Habitat-enriched fish were faster at finding their way out of a four-armed maze than individuals reared in standard, non-enriched hatchery conditions. The enrichment promoted neural plasticity in the telencephalon with a positive effect on later life. Cognitive abilities can also be improved by exposure to environmental change during early ontogeny (Kotrschal & Taborsky, 2010). Individuals of *S. pleurospilus* experiencing changed food rations early in life outperformed fish kept on constant rations in a later learning task. This difference in learning abilities between individuals remained constant between juvenile and adult stages of the same fish tested 1 year apart.

Environmental complexity and different kinds of sensory input can lead to neural developmental and plastic modifications; but if understimulated, fishes typically develop smaller brains (Mayer et al., 2011). In salmonids, tank-rearing influences anatomical characteristics such as the development of the forebrain (telencephalon), cerebellum and optic tectum (Marchetti & Nevitt, 2003; Kihslinger & Nevitt, 2006; Kihslinger et al., 2006). Cultured Oncorhynchus spp. have smaller brains, dendrite modifications of the dendritic structure and gene expression of neuroplasticity markers. Also, sensory organs, such as the lateral line system and eyes, may be modified and influence behavioural performance of hatchery fishes (Marchetti & Nevitt, 2003; Anras & Lagardere, 2004). Also, the brain gene expression profiles in S. salar are affected by rearing environments, such as hatchery and river, as well as between reproductive tactics independent of rearing environment (Aubin-Horth et al., 2005). Important in the growth of the various regions of the brain are proliferation and incorporation of new neurons that handle new information. In D. rerio, habitat enrichment during early rearing increased proliferating cell nuclear antigen positive cells in the forebrain (von Krogh et al., 2010), and in the weakly electric fish Brachyhypopomus gauderio Giora & Malabarba 2009 from a natural environment had greater rates of brain cell proliferation than conspecifics reared in social or isolated captive environments (Dunlap *et al.*, 2011).

#### Mate choice

Early experience may influence mate choice. In cross-fostering experiments with the closely related cichlid species pair *Haplochromis pundamilia* (Seehausen & Bouton 1998) and *Haplochromis nyererei* Witte-Maas & Witte 1985 of Lake Victoria, Verzijden & Cate (2007) showed that young females develop a sexual preference for males of their foster mothers' species. Crosses were made both ways and showed that these species reversed their species assortative mating preferences when developed in the mouth of a female of the other species. It is not known whether the colour or smell of the foster mother were imprinted in the offspring, but obviously, this learning may be important in the speciation process of the haplochromine cichlids of the species flocks in the great African lakes.

Thus, a whole suite of behavioural reactions can be influenced by early experiences. Since much of this research is very recent, it is expected that several more examples will be published rapidly enhancing knowledge about behavioural effects of early experiences.

#### **FUTURE RESEARCH**

Important future questions are to what degree parental and early environmental influences affect future performances of fishes and what the genetic, epigenetic, neural and hormonal mechanisms are. For instance, environmental influences during embryogenesis may have effects on the adult phenotype that are difficult to realize when monitoring performances of the organism in the field. Without this knowledge, present interpretation of, for example, climate change effects may be skewed or wrong. Because of the time lapse between cause and effect, effects of such early influences are chiefly discovered through long-lasting experimental research.

The links between early influences, behaviour, growth and life-history characteristics are still poorly developed, and new knowledge about these relationships will be valuable for understanding of population and species effects in relation to natural and anthropogenic habitat modifications and climate change (Fig. 1). Future research may focus on genetic mechanisms influencing phenotypic plasticity caused by early and trans-generational influences, and link effects of early influences and later norms of reaction in life-history and behavioural traits. Arising questions are: (1) Is there a relationship between boldness, the early behavioural syndrome linking activity to aggressiveness and exploration tendency (Andriaenssens & Johnsson, 2013), migratory tendency and epigenetics? (2) Does embryonic temperature or other environmental stimuli influence later activity, boldness and social behaviour? (3) Is the tendency to migrate in partly migratory populations influenced by epigenetics? (4) Are other life-history characteristics influenced by environmental stimuli during embryogenesis? (5) To what degree is the phenology of species influenced by epigenetics? (6) To what degree is later phenotypic response to early environmental influences inherited and thereby influenced by natural selection (Walsh & Reznick, 2011)? (7) How is individual fitness related to this phenotypic variability? (8) Is there a cost of phenotypic

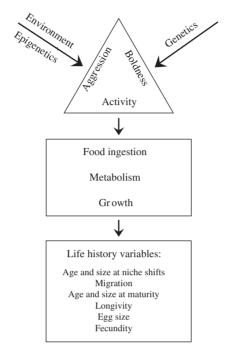


Fig. 1. Genetics and environment affect personality development, social behaviour and activity of fishes. The environmental effect may be partly epigenetic with likely effects on food consumption, metabolic rate and amount of surplus energy used for growth with subsequent effects on life-history traits.

flexibility? (9) Which epigenetic mechanisms are involved in phenotypically plastic reaction norms? (10) How are epigenetic effects conveyed to later activated life stages (neural, hormonal, enzymes)?

Moran & Pérez-Figueroa (2011) compare methylation patterns between mature and immature parr of S. salar from two different rivers in order to infer if DNA methylation differences may be related to their maturation condition. Contrary to mature parr, immature parr migrate to the ocean and become large fish before they attain maturity. Their results revealed that genetically similar mature and immature salmon parr present high levels of DNA methylation variation in two of the three analysed tissues (liver, brain and gonads), and hypothesized that early maturation may be mediated mostly by epigenetic processes rather than by genetic differences between the young fish as suggested by Piché et al. (2008). If there is an epigenetic relationship between migratory tendency and environmental influence during embryogenesis, this may explain the finding of Giger et al. (2006) that there is no apparent inherited genetic difference between resident and migratory S. trutta, but a difference exists in their gene expressions. Life-history characteristics are quantitative traits influenced by a large number of genes, whereas the expression may be determined by a few control genes under strong environmental influence. Also, there may be density-dependent influence conveyed during early juvenile stages related to growth, metabolic rate or degree of lipid reserve energy at that time (Jonsson & Jonsson, 1993; Olsson et al., 2006), but neither epigenetic nor density-dependent influences on migration in partly migratory populations have been studied very much.

Early environmental effects on later performance in fishes are as yet a poorly developed field. New studies should be aimed at exploiting the mechanisms of this plasticity, and how the norms of reaction are and reflect differences in ability of the fishes to respond to environmental change. By increasing the understanding of interactions between environments and genotypes, the capacity to predict population responses to early influences on later phenotypic plasticity will be refined. Evidence indicates that the degree of plasticity is heritable and that population differences reflect adaptive responses formed by natural selection to local environments. In evolutionary ecology and conservation biology, it is important to understand how these vary within populations across environmental gradients, among populations and among species. Such knowledge may influence evaluation of ecological responses observed across environmental gradients.

Financially, this review was supported by the Norwegian Research Council's programme Norklima (Project no 225014/E10) and the Norwegian Institute for Nature Research. F. Segers and an anonymous referee gave helpful comments to an earlier version of the paper.

#### References

- Abozaid, H., Wessels, S. & Hörstgen-Schwark, G. (2012). Elevated temperature applied during gonadal transformation leads to male bias in zebrafish (*Danio rerio*). *Sexual Development* **6**, 201–209. doi: 10.1159/000336297
- Ali, M., Nicieza, A. & Wootton, R. J. (2003). Compensatory growth in fishes: a response to growth depression. *Fish and Fisheries* **4**, 147–190.
- Alm, G. (1959). Connection between maturity, size and age in fishes. Report of the Institute of Freshwater Research Drottningholm 40, 5–145.
- Álvarez, D. & Nicieza, G.A. (2005). Compensatory growth 'defends' energy levels but not growth trajectories in brown trout, *Salmo trutta* L. *Proceedings of the Royal Society B* **272**, 601–607. doi: 10.1098/rspb.2004.2991
- Álvarez, D., Cano, J. M. & Nicieza, A. G. (2006). Microgeographic variation in metabolic rate and energy storage of brown trout: countergradient selection or thermal sensitivity? *Evolutionary Ecology* **20**, 345–363. doi: 10.1007/s10682-006-0004-1
- Andriaenssens, B. & Johnsson, J. (2013). Natural selection, plasticity and the emergence of a behavioural syndrome in the wild. *Ecology Letters* **16**, 47–55. doi: 10.1111/ele.12011
- Angers, B., Castonguay, E. & Massicotte, R. (2010). Environmentally induced phenotypes and DNA methylation: how to deal with unpredictable conditions until the next generation and after. *Molecular Ecology* **19**, 1283–1295. doi: 10.1111/j.1365-294X.2010.04580.x
- Angilletta, M. J., Niewiarowski, P. H. & Navas, C. A. (2002). The evolution of thermal physiology in ectotherms. *Journal of Thermal Biology* 27, 249–268. doi: 10.1016/S0306-4565(01)00094-8
- Anras, M. L. B. & Lagardere, J. P. (2004). Domestication et comportement chez les poisons téléostéens. *INRA Productions Animales* 17, 211–215.
- Arendt, J. D. (1997). Adaptive intrinsic growth-rates: an integration across taxa. *Quarterly Review of Biology* **72**, 149–177.
- Arendt, J. D. (2000). Allocation of cells to proliferation vs. differentiation and its consequences for growth and development. *Journal of Experimental Zoology* **288**, 219–234. doi: 10.1002/1097-010X(20001015)288:3<219::AID-JEZ3>3.0.CO;2-C
- Arnold, C. & Taborsky, B. (2010). Social experience in early ontogeny has lasting effects on social skills in cooperatively breeding cichlids. *Animal Behaviour* **79**, 621–630. doi: 10.1016/j.anbehav.2009.12.008
- Aubin-Horth, N., Letcher, B. H. & Hoffmann, H. A. (2005). Interaction of rearing environment and reproductive tactic on gene expression profiles in Atlantic salmon. *Journal of Heredity* **96**, 261–278. doi: 10.1093/jhered/esi030

- Azuma, T., Takeda, K., Doi, T., Muto, K., Akutsu, M., Sawada, M. & Adachi, S. (2004). The influence of temperature on sex determination in sockeye salmon *Oncorhynchus nerka*. *Aquaculture* **234**, 461–473. doi: 10.1016/j.aquaculture.2003.11.023
- Baroiller, J. F. & D'Cotta, H. (2001). Environment and sex determination in farmed fish. *Comparative Biochemistry and Physiology* **130**, 399–409.
- Baroiller, J. F., D'Cotta, H. & Saillant, E. (2009a). Environmental effects of fish sex determination and differentiation. *Sexual Development* 3, 118–135. doi: 10.1159/000223077
- Baroiller, J. F., D'Cotta, H., Bezault, E., Wessels, S. & Hoertigen-Schwark, G. (2009b). Tilapia sex determination: where temperature and genetics meet. *Comparative Biochemistry and Physiology A* **153**, 30–38. doi: 10.1016/j.cbpa.2008.11.018
- Beamish, F. W. H. (1993). Environmental sex determination in southern brook lamprey, *Ichthyomyzon gagei*. Canadian Journal of Fisheries and Aquatic Sciences **50**, 1299–1307. doi: 10.1139/f93-147
- Bell, A. M., Dingemanse, N. J., Hankison, S. J., Langenhof, M. B. W. & Rollins, K. (2011). Early exposure to nonlethal predation risk by size-selective predators increases somatic growth and decreases size at adulthood in threespined sticklebacks. *Journal of Evolutionary Biology* 24, 943–954. doi: 10.1111/j.1420-9101.2011.02247.x
- Berejikian, B. A., Tezak, E. P., Riley, S. C. & LaRae, A. L. (2001). Competitive ability and social behaviour of juvenile steelhead reared in enriched and conventional hatchery tanks and a stream environment. *Journal of Fish Biology* **59**, 1600–1613. doi: 10.1111/j.1095-8649.2001.tb00224.x
- Berglund, I. (1992). Growth and early sexual maturation in Baltic salmon (*Salmo salar*) parr. *Canadian Journal of Zoology* **70**, 205–211. doi: 10.1139/z92-032
- Bird, A. (2002). DNA methylation patterns and epigenetic memory. *Genes and Development* **16,** 6–21. doi: 10.1101/gad.947102
- Biro, P. A. & Stamps, J. A. (2008). Are animal personality traits linked to life history productivity? *Trends in Ecology and Evolution* **23**, 361–368. doi: 10.1016/j.tree.2008.04.003
- Blazquez, M. & Somoza, G. M. (2010). Fish with thermolabile sex determination (TSD) as models to study brain sex differentiation. *General and Comparative Endocrinology* **166**, 470–477. doi: 10.1016/j.ygcen.2009.10.004
- de Boo, H. A. & Harding, J. E. (2006). The developmental origins of adult disease (Barker) hypothesis. *Australian and New Zealand Journal of Obstetrics and Gynaecology* **46**, 4–14.
- Bossdorf, O., Richards, C. L. & Pigliucci, M. (2008). Epigenetics for ecologists. *Ecology Letters* **11,** 106–115.
- Bownds, C., Wilson, R. & Marshall, D. J. (2010). Why do colder mothers produce larger eggs? An optimality approach. *Journal of Experimental Biology* **213**, 3796–3801. doi: 10.1242/jeb.043356
- Braithwaite, V. A. & Salvanes, A. G. V. (2005). Environmental variability in the early rearing environment generates behaviourally flexible cod: implications for rehabilitating wild populations. *Proceedings of the Royal Society B* **272**, 1107–1113. doi: 10.1098/rspb.2005.3062
- Brett, J. R. (1952). Temperature tolerance in young Pacific salmon, genus *Oncorhynchus*. *Journal of the Fisheries Research Board of Canada* **9**, 265–323.
- Brockmark, S. & Johnsson, J. I. (2010). Reduced hatchery rearing density increases social dominance, post release growth, and survival in brown trout (*Salmo trutta*). *Canadian Journal of Fisheries and Aquatic Sciences* **67**, 288–295. doi: 10.1139/F09-185
- Brown, C., Davidson, T. & Laland, K. (2003). Environmental enrichment and prior experience of live prey improve foraging behaviour in hatchery-reared Atlantic salmon. *Journal of Fish Biology* **63** (Suppl. A), 187–196. doi: 10.1111/j.1095-8649.2003.00208.x
- Brown, C., Jones, F. & Braithwaite, V. A. (2007). Correlation between boldness and body mass in natural populations of poecilid Brachyrhaphis episcopi. *Journal of Fish Biology* **71**, 1590–1601. doi: 10.1111/j.1095-8649.2007.01627.x
- Bunnell, D. B., Thomas, S. E. & Stein, R. A. (2007). Prey resources before spawning influence gonadal investment of female, but not male, white crappie. *Journal of Fish Biology* **70**, 1838–1854. doi: 10.1111/j.1095-8649.2007.01459.x

- Burton, T., Killen, S. S., Armstrong, J. D. & Metcalfe, N. D. (2011). What causes intraspecific variation in resting metabolic rate and what are its ecological consequences? *Proceedings of the Royal Society B* **278**, 3465–3473. doi: 10.1098/rspb.2011.1778
- Burton, T., McKelvey, S., Stewart, D. C., Armstrong, J. D. & Metcalfe, N. B. (2013). Offspring investment in wild Atlantic salmon (*Salmo salar*): relationships with smolt age and spawning condition. *Ecology of Freshwater Fish* **22**, 317–321. doi: 10.1111/eff.12019
- Cabej, N. R. (2012). Epigenetic Principles of Evolution. London: Elsevier.
- Campos, C., Valente, L. M. P., Conceicao, L. E. C., Engrola, S., Sousa, V., Rocha, E. & Fernandes, J. M. O. (2013). Incubation temperatures induces changes in muscle cellularity and gene expression in Senegalese sole (*Solea senegalensis*). *Gene* 516, 209–217. doi: 10.1016/j.gene.2012.12.074
- Carere, C., Drent, P. J., Koolhaas, J. M. & Groothuis, T. G. G. (2005). Epigenetic effects on personality traits: early food provisioning and sibling competition. *Behaviour* **142**, 1329–1355. doi: 10.1163/156853905774539328
- Champagne, F.A. (2008). Epigenetic mechanisms and the transgenerational effects of maternal care. *Frontiers in Neuroendocrinology* **29**, 386–397. doi: 10.1016/j.yfrne.2008.03.003
- Chandler, V. L. (2007). Paramutation: from maize to mice. *Cell* **128**, 641–645. doi: 10.1016/j.cell.2007.02.007
- Chapman, B., Ward, A. J. W. & Krause, J. (2008). Schooling and learning: early social environment predicts social learning ability in guppy. *Animal Behaviour* **76**, 923–929. doi: 10.1016/j.anbehav.2008.03.022
- Chapman, B., Morrell, L. J. & Krause, J. (2010). Unpredictability in food supply during early life influences boldness in fish. *Behavioral Ecology* **21**, 501–506. doi: 10.1093/beheco/arq003
- Charnov, E.L. & Bull, J. (1977). When is sex environmentally determined? *Nature* **266**, 828–830. doi: 10.1038/266828a0
- Chik, F., Szyf, M. & Rabbani, S. A. (2011). Role of epigenetics in cancer initiation and progression. *Advances in Experimental Medicine and Biology* **720,** 91–104. doi: 10.1007/978-1-4614-0254-1\_8
- Clutton-Brock, T. H. (1991). *Evolution of Parental Care*. Princeton, NJ: Princeton University Press
- Conover, D. O. (1984). Adaptive significance of temperature-dependent environments and genetic sex determination in fish. *American Naturalist* **123**, 297–313.
- Conover, D. O. & Heins, S. W. (1987). Adaptive variation in environmental and genetic sex determination in a fish. *Nature* **326**, 496–498. doi: 10.1038/326496a0
- Conover, D. O., Van Vorhees, D. A. & Ehtisham, A. (1992). Sex ratio selection and the evolution of environmental sex determination in laboratory populations of *Menidia menidia*. *Evolution* **46**, 1722–1730.
- Craig, J. K., Foote, C. J. & Wood, C. C. (1996). Evidence for temperature-dependent sex determination in sockeye salmon (*Oncorhynchus nerka*). *Canadian Journal of Fisheries and Aquatic Sciences* **53**, 141–147. doi: 10.1139/f95-152
- Crews, D. (2010). Epigenetics, brain, behavior, and the environment. *Hormones* 9, 41–50.
- Danchin, E. & Wagner, R. H. (2010). Inclusive heritability: combining genetic and non-genetic information to study animal behaviour and culture. *Oikos* **119**, 210–218.
- Davey, A. J. H. & Jellyman, D. J. (2005). Sex determination in freshwater eels and management options for manipulation of sex. *Reviews in Fish Biology and Fisheries* **15**, 37–52. doi: 10.1007/s11160-005-7431-x
- Dawson, M. A. & Kouzarides, T. (2012). Cancer epigenetics: from mechanism to therapy. *Cell* **150**, 12–27. doi: 10.1016/j.cell.2012.06.013
- Day, T. & Rowe, L. (2002). Developmental thresholds and the evolution of reaction norms for age and size at life-history transitions. *American Naturalist* **159**, 338–350. doi: 10.1086/338989
- Desai, M. & Hales, C. N. (1997). Role of fetal and infant growth in programming metabolism in later life. *Biological Reviews* **72**, 329–348. doi: 10.1017/s0006323196005026
- Descamps, S., Boutin, S., Berteaux, D., McAdam, A. G. & Gaillard, J. M. (2008). Cohort effects in red squirrels: the influence of density, food abundance and temperature on future survival and reproductive success. *Journal of Animal Ecology* **77**, 305–314. doi: 10.1111/j.1365-2656.2007.01340.x

- Dittman, A. H. & Quinn, T. P. (1996). Homing in Pacific salmon. Mechanisms and ecological basis. *Journal of Experimental Biology* **199**, 83–91.
- Dobson, S. H. & Holmes, R. M. (1984). Compensatory growth in rainbow trout, *Salmo gaird-neri* Richardson. *Journal of Fish Biology* **25**, 649–656. doi: 10.1111/j.1095-8649.1984. tb04911.x
- Donelson, J. M., McCormick, M. I. & Munday, P. H. (2008). Parental condition affects early life-history of a coral reef fish. *Journal of Experimental Marine Biology and Ecology* **360**, 109–116. doi: 10.1016/j.jembe.2008.04.007
- Dunlap, K. D., Silva, A. C. & Chung, M. (2011). Environmental complexity, seasonality and brain cell proliferation in a weakly electric fish, *Brachyhypopomus gauderio*. *Journal of Experimental Biology* 214, 794–805. doi: 10.1242/jeb.051037
- Ebbeson, L. E. & Braithwaite, V. A. (2012). Environmental effects on fish neural plasticity and cognition. *Journal of Fish Biology* **81,** 2151–2174. doi: 10.1111/j.1095-8649.2012. 03486.x
- Einum, S. & Fleming, I. A. (2004). Environmental unpredictability and offspring size: conservative versus diversified bet-hedging. *Evolutionary Ecology Research* **2004**, 443–455.
- Elliott, J. M. (1994). *Quantitative Biology and the Brown Trout*. Oxford: Oxford University Press.
- Eriksen, M. S., Faerevik, G., Kittilsen, S., McCormick, M. I., Damsgård, B., Braithwaite, V. A., Braastad, B. O. & Bakken, M. (2011). Stressed mothers troubled offspring: a study of behavioural maternal effects in farmed *Salmo salar*. *Journal of Fish Biology* **79**, 575–586. doi: 10.1111/j.1095-8649.2011.03036.x
- Fernandino, J. I., Hattori, R. S., Kishii, A., Strussmann, C. A. & Somoza, G. M. (2012). The cortisol and androgen pathway cross talk in high temperature-induced masculinization: the 11 beta-hydroxysteroid dehydrogenase as a key enzyme. *Endocrinology* **153**, 6003–6011. doi: 10.1210/en.2012-1517
- Finstad, A. G. & Jonsson, B. (2012). Effect of incubation temperature on growth performance in Atlantic salmon. *Marine Ecology Progress Series* **454**, 75–82. doi: 10.3354/meps09643
- Fleming, I. A. & Gross, M. R. (1990). Latitudinal clines: a trade-off between egg number and size in Pacific salmon. *Ecology* **71**, 1–11.
- Fleming, I. A., Jonsson, B., Gross, M. R. & Lamberg, A. (1996). An experimental study of the reproductive behaviour and success of farmed and on wild Atlantic salmon (*Salmo salar*). *Journal of Applied Ecology* **33**, 893–905.
- Fleming, I. A., Lamberg, A. & Jonsson, B. (1997). Effects of early experience on the reproductive performance of Atlantic salmon. *Behavioral Ecology* **8**, 470–480. doi: 10.1093/beheco/8.5.470
- Forseth, T., Ugedal, O. & Jonsson, B. (1995). Inter and intra cohort variation in the life history of Arctic charr (*Salvelinus alpinus*). *Nordic Journal of Freshwater Research* **71**, 237–244.
- Forseth, T., Næsje, T. F., Jonsson, B. & Hårsaker, K. (1999). Juvenile migration in brown trout: a consequence of energetic state. *Journal of Animal Ecology* **68**, 783–793. doi: 10.1046/j.1365-2656.1999.00329.x
- Fox, C. W., Waddell, K. J. & Mousseau, T. A. (1995). Parental host plant affects offspring life histories in a seed beetle. *Ecology* **76**, 402–411.
- Francis, R. C. & Barlow, G. W. (1993). Social control of primary sex differentiation in the Midas cichlid. *Proceedings of the National Academy of Sciences of the United States of America* **90,** 10673–10675.
- Gabillard, J. C., Weil, C., Rescan, P. Y., Navarro, I., Gutierrez, J. & Le Bail, Y. P. (2005). Does the GH/IGF system mediate the effect of water temperature on fish growth? A review. *Cybium* **29**, 107–117.
- Giesel, J. T. (1986). Effects of parental photoperiod regime on progeny development time in *Drosophila simulans*. *Evolution* **40**, 649–651.
- Giesel, J. T. (1988). Effects of parental photoperiod on development time and density sensitivity of progeny in *Drosophila melanogaster*. *Evolution* **42**, 1348–1350.
- Giesing, E. R., Suski, C. D., Warner, R. E. & Bell, A. M. (2011). Female sticklebacks transfer information via eggs: effects of maternal experience with predators on offspring. *Pro*ceedings of the Royal Society B 278, 1753–1759. doi: 10.1098/rspb.2010.1819

- Giger, T., Excoffier, L., Day, P. J. R., Champigneulle, A., Hansen, M. M., Powell, R. & Largiader, C. R. (2006). Life histories shapes gene expression in salmonids. *Current Biology* 16, 281–282.
- Gilbert, S. F. (2005). Mechanisms for the environmental regulation of gene expression: ecological aspects of animal development. *Journal of Biosciences* **30**, 65–74.
- Gilbert, S. F. & Epel, D. (2008). *Ecological Developmental Biology*. Sunderland, MA: Sinauer Associates, Inc.
- Gjerde, B. (1984). Response to individual selection for age at sexual maturity in Atlantic salmon. *Aquaculture* **38**, 229–240. doi: 10.1016/0044-8486(84)90147-9
- Goodey, W. & Liley, N. R. (1986). The influence of early experience on escape behaviour in the guppy (*Poecilia reticulata*). *Canadian Journal of Zoology* **64**, 885–888. doi: 10.1139/z86-132
- Gräff, J., Kim, D., Dobbin, M.M. & Tsai, L.-H. (2011). Epigenetic regulation of gene expression in physiological and pathological brain processes. *Physiological Reviews* **91**, 603–649. doi: 10.1152/physrev.00012.2010
- Green, B. S. & McCormick, M. I. (2005). Maternal and paternal effects determine size, growth and performance in larvae of a tropical reef fish. *Marine Ecology Progress Series* **289**, 263–272.
- Gregersen, F., Vøllestad, L. A., Østbye, K., Aass, P. & Hegge, O. (2011). Temperature and food-level effects on reproductive investment and egg mass in vendace, *Coregonus albula. Fisheries Management and Ecology* **18,** 263–269. doi: 10.1111/j.1365-2400.2010.00779x
- Groothius, T.G.G., Müller, W., von Engelhardt, N., Carere, C. & Eising, C. (2005). Maternal hormones as a tool to adjust offspring phenotype in avian species. *Neuroscience and Biobehavioral Reviews* **29**, 329–352. doi: 10.1016/j.neubiorev.2004.12.002
- Gudsnuk, K. M. & Champagne, F. A. (2011). Epigenetic effects of early developmental experiences. *Clinics in Perinatology* **38**, 703–717.
- Gurney, W. S. C., Jones, W., Veitch, A. R. & Nisbet, R. M. (2003). Resource allocation, hyperphagia and compensatory growth in juveniles. *Ecology* **84**, 2777–2787. doi: 10.1890/02-0536
- Hansen, L. P., Jonsson, N. & Jonsson, B. (1993). Oceanic migration of homing Atlantic salmon. *Animal Behaviour* **45**, 927–941.
- Harrald, M., Neat, F. C., Wright, P. J. & Huntingford, F. A. (2010). Differences in thermal growth responses of juvenile Atlantic cod (*Gadus morhua* L.) from the east and west coast of Scotland. *Environmental Biology of Fishes* 87, 187–194. doi: 10.1007/s10641-010-9586-0
- Harrison, X. A., Blount, J. D., Inger, R., Norris, D. R. & Bearhop, S. (2010). Carry-over effects as drivers of fitness differences in animals. *Journal of Animal Ecology* **80**, 4–18. doi: 10.1111/j.1365-2656.2010.01740.x
- Hasler, A. D. & Scholz, A. T. (1983). Olfactory Imprinting and Homing in Salmon. Investigations into the Mechanism of the Imprinting Process. Berlin: Springer Verlag.
- Hasyashi, Y., Kobira, H., Yamaguchi, T., Shiraishi, E., Yazawa, T., Hirai, T., Kamei, Y. & Kitano, T. (2010). High temperature causes masculinization of genetically female medaka by elevation of cortisol. *Molecular Reproduction and Development* 77, 679–686. doi: 10.1002/mrd.21203:
- Heath, D. D. & Blouw, D. M. (1998). Are maternal effects in fish adaptive or merely physiological side effects? In *Maternal Effects as Adaptations* (Mousseau, T. A. & Fox, C. W., eds), pp. 178–201. New York, NY: Oxford University Press.
- Heath, D. D., Fox, C. W. & Heath, J. W. (1999). Maternal effects on offspring size: variation through early development of Chinook salmon. *Evolution* **53**, 1605–1611.
- Heiligenberg, W. (1965). Color polymorphism in the males of an African cichlid fish. *Journal of Zoology* **146**, 95–97. doi: 10.1111/j.1469-7998.1965.tb05202.x
- Holbrook, R. I. & Burt de Perera, T. (2011). Fish navigation in the vertical dimension: can fish use hydrostatic pressure to determine depth? *Fish and Fisheries* **12**, 370–379. doi: 10.1111/j.1467-2979.2010.00399.x
- Holliday, R. & Pugh, J. E. (1975). DNA modification mechanisms and gene activity during development. *Science* **187**, 226–232. doi: 10.1126/science.187.4173.226

- Hollis, K. L., Langworthy-Lam, K. S., Blouin, L. A. & Romano, M. C. (2004). Novel strategies of subordinate fish competing for food: learning when to fold. *Animal Behaviour* 68, 1155–1164. doi: 10.1016/j.anbehav.2003.11.022
- Huntingford, F. A., Wright, P. J. & Tierney, J. F. (1994). Adaptive variation in antipredator behaviours in threespine stickleback. In *The Evolutionary Biology of Threespine Stickle-back* (Bell, M. A. & Foster, S. A., eds), pp. 345–380. Oxford: Oxford University Press.
- Hyvarinen, P. & Rodewald, P. (2013). Enriched rearing improves survival of hatchery-reared Atlantic salmon smolts during migration in the River Tornionjoki. *Canadian Journal of Aquatic Sciences* **70**, 1386–1395. doi: 10.1139/cjfas-2013-0147
- Iguchi, K. (2012). Larger eggs at lower water temperature as a measure to assure effective hatchling size in the landlocked form of Ayu *Plecoglossus altivelis*. *Ichthyological Research* **59,** 20–25. doi: 10.1007/s10228-011-0251-1
- Iida, A., Shimada, A., Shima, A., Takamatsu, N., Hori, H., Takeuchi, K. & Koga, A. (2006). Targeted reduction of the DNA methylation level with 5-azacytidine promotes excision of the medaka Fish Tol2 transposable element. Genetics Research (Cambridge) 87, 187–193.
- Inness, C. L. W. & Metcalfe, N. B. (2008). The impact of dietary restriction, intermittent feeding and compensatory growth on reproductive investment and lifespan in a short-lived fish. *Proceedings of the Royal Society B* **275**, 1703–1708. doi: 10.1098/rspb.2008.0357
- Jablonka, E. (2001). The systems of inheritance. In *Cycles of Contingency* (Óyama, S., Griffith, P. & Grey, R., eds), pp. 99–116. Cambridge, MA: MIT Press.
- Jablonka, E. & Raz, G. (2009). Transgenerational epigenetic inheritance: prevalence, mechanisms, and implications for the study of heredity and evolution. *Quarterly Review of Biology* 84, 131–176. doi: 10.1086/598822
- Janzen, F.J. (1994). Climate change and temperature-dependent sex determination in reptiles. *Proceedings of the National Academy of Sciences* **91,** 7487–7490.
- Johnsson, J.I. & Bohlin, T. (2005). Compensatory growth for free? A field experiment on brown trout, *Salmo trutta*. *Oikos* **111**, 31–38. doi: 10.1111/j.0030-1299.2005.13972.x
- Johnston, I. A. (2006). Environment and plasticity of myogenesis in teleost fish. *Journal of Experimental Biology* **209**, 2249–2264. doi: 10.1111/j.1420-9101.2011.02247.x
- Jones, P. A. & Taylor, S. M. (1980). Cellular differentiation, cytidine analogs, and DNA methylation. Cell 20, 85–93. doi: 10.1016/0092-8674(80)90237-8
- Jonsson, B. (1985). Life history patterns of freshwater resident and sea-run migrant brown trout in Norway. *Transactions of the American Fisheries Society* **114,** 182–194. doi: 10.1577/1548-8659(1985)114<182:LHPOFR>2.0.CO;2
- Jonsson, B. & Hindar, K. (1982). Reproductive strategy of dwarf and normal Arctic charr (Salvelinus alpinus) from Vangsvatnet Lake, western Norway. Canadian Journal of Fisheries and Aquatic Sciences 39, 1404–1413. doi: 10.1139/f82-189
- Jonsson, B. & Jonsson, N. (1993). Partial migration: niche shift versus sexual maturation in fishes. *Reviews in Fish Biology and Fisheries* **3**, 348–365.
- Jonsson, N. & Jonsson, B. (1999). Trade-off between egg size and numbers in brown trout. *Journal of Fish Biology* **55**, 767–783. doi: 10.1006/jfbi.1999.1035
- Jonsson, N. & Jonsson, B. (2003). Energy density and content of Atlantic salmon: variation among developmental stages and types of spawners. *Canadian Journal of Fisheries and Aquatic Sciences* **60**, 506–516. doi: 10.1139/f03-042
- Jonsson, B. & Jonsson, N. (2006). Life history effects of migratory costs in anadromous brown trout *Salmo trutta*. *Journal of Fish Biology* **69,** 860–869. doi: 10.1111/j.1095-8649. 2006.01160.x
- Jonsson, B. & Jonsson, N. (2011). *Ecology of Atlantic Salmon and Brown Trout: Habitat as a Template for Life Histories*. Dordrecht: Springer.
- Jonsson, B., Jonsson, N. & Hansen, L. P. (1990a). Does juvenile experience affect migration and spawning of adult Atlantic salmon. *Behavioral Ecology and Sociobiology* **26**, 225–230.
- Jonsson, N., Jonsson, B. & Hansen, L. P. (1990b). Partial segregation in the timing of migration of Atlantic salmon of different ages. *Animal Behaviour* **40**, 313–321.
- Jonsson, N., Jonsson, B. & Fleming, I. A. (1996). Does early growth rate cause a phenotypically plastic response in egg production of Atlantic salmon? *Functional Ecology* **10**, 89–96.
- Jonsson, N., Næsje, T. F., Jonsson, B., Saksgård, R. & Sandlund, O. T. (1999). The influence of piscivory on life history traits of brown trout. *Journal of Fish Biology* 55, 1129–1141. doi: 10.1111/j.1095-8649.1999.tb02064.x

- Jonsson, N., Jonsson, B. & Hansen, L. P. (2005). Does climate during embryonic development influences parr growth and age of seaward migration in Atlantic salmon (*Salmo salar*) smolts? *Canadian Journal of Fisheries and Aquatic Sciences* **62**, 2502–2508.
- Jonsson, B., Finstad, A. G. & Jonsson, N. (2012). Winter temperature and food quality affect age and size at maturity in ectotherms: an experimental test with Atlantic salmon. *Canadian Journal of Fisheries and Aquatic Sciences* 69, 1817–1826.
- Jonsson, B., Jonsson, N. & Finstad, A. G. (2013). Effects of temperature and food quality on age at maturity of ectotherms: an experimental test of Atlantic salmon. *Journal of Animal Ecology* **82**, 201–210. doi: 10.1111/j.1365-2656.2012.02022.x
- Jump, D. B. & Clark, S. D. (1999). Regulation of gene expression by dietary fat. *Annual Review of Nutrition* **19**, 63–90. doi: 10.1146/annurev.nutr.19.1.63
- Kaiser, S. & Sachser, N. (2009). Effects of prenatal social stress on offspring development: pathology or adaptation? *Current Directions in Psychological Science* **18**, 118–121. doi: 10.1111/j.1467-8721.2009.01620.x
- Kakutani, T. (2002). Epi-alleles in plants: inheritance of epigenetic information over generations. *Plant Cell Physiology* **43**, 1106–1111. doi: 10.1093/pcp/pcf131
- Kennedy, J., Witthams, P.R., Nash, R.D.M. & Fox, C.J. (2008). Is fecundity in plaice (*Pleuronectes platessa* L.) down-regulated in response to reduced food intake during autumn? *Journal of Fish Biology* **72**, 78–92. doi: 10.1111/j.1095-8649.2007.01651.x
- Kerr, L. A. & Secor, D. H. (2010). Latent effects of early life history on partial migration for an estuarine-dependent fish. *Environmental Biology of Fishes* **89**, 479–492.
- Khrameeva, E. E., Mironov, A. A., Fedonin, G. G., Khaitovich, P. & Gelfand, M. S. (2012). Spatial proximity and similarity of the epigenetic state of genome domains. *PLoS One* **7**, e33947. doi: 10.1371/journal.pone.0033947
- Kihslinger, R. L. & Nevitt, G. A. (2006). Early rearing environment impacts cerebellar growth in juvenile salmon. *Journal of Experimental Biology* **209**, 504–509. doi: 10.1242/jeb.02019
- Kihslinger, R. L., Lema, S. C. & Nevitt, G. A. (2006). Environmental rearing conditions produce forebrain differences in wild Chinook salmon *Oncorhynchus tshawytscha*. *Comparative Biochemistry and Physiology A* **145**, 145–151. doi: 10.1016/j.cbpa.2006.06.041
- Killen, S. S., Marras, S., Ryan, M. R., Domenici, P. & McKenzie, D. J. (2012). A relationship between metabolic rate and risk-taking behaviour is revealed during hypoxia in juvenile European sea bass. *Functional Ecology* 26, 134–143. doi: 10.1111/j.1365-2435.2011.01920.x
- Knudson, E. I. (2004). Sensitive periods in the development of the brain and behavior. *Journal of Cognitive Neuroscience* **16**, 1412–1425. doi: 10.1162/0898929042304796
- Koops, M., Hutchings, J. A. & Adams, B. K. (2003). Environmental predictability and the cost of imperfect information: influences on offspring size variability. *Evolutionary Ecology Research* 5, 29–42.
- Korwin-Kossakowski, M. (2008). The influence of temperature during the embryonic period on larval development and growth in carp, *Cyprinus carpio* L., and grass carp, *Ctenopharyngodon idella* (Val.): theoretical and practical aspects. *Archives of Polish Fisheries* **16**, 231–314. doi: 10.2478/s10086-008-0020-6
- Kotrschal, A. & Taborsky, B. (2010). Environmental change enhances cognitive abilities in fish. *PLoS Biology* **8**, e1000351. doi: 10.1371/journal.pbio.1000351
- Kotrschal, A., Heckel, G., Bonfils, D. & Taborsky, B. (2012). Life-stage specific environments for inducible maternal effects. *Evolutionary Ecology* 26, 123–137. doi: 10.1007/s10682-011-9495-5
- Koumoundouros, G., Ashton, C., Sfakianakis, D. G., Divanach, P., Kentouri, M., Anthwal, N. & Stickland, N. C. (2009). Thermally induced phenotypic plasticity of swimming performance in European sea bass *Dicentrarchus labrax* juveniles. *Journal of Fish Biology* 74, 1309–1322. doi: 10.1111/j.1095-8649.2009.02206.x
- von Krogh, K., Sørensen, C., Nilsson, G. E. & Øverli, Ø. (2010). Forebrain cell proliferation, behavior, and physiology of zebrafish, *Danio rerio*, kept in enriched or barren environments. *Physiology and Behavior* **101**, 32–39. doi: 10.1016/j.physbeh.2010.04.003
- Krueger, W. H. & Oliveira, K. (1999). Evidence for environmental sex determination in the American eel, *Anguilla rostrata*. *Environmental Biology of Fishes* **55**, 381–389. doi: 10.1023/A:1007575600789

- Kutschera, U. & Niklas, K. J. (2004). The modern theory of biological evolution: an expanded synthesis. *Naturwissenschaften* **91**, 255–276. doi: 10.1007/s00114-004-01515-y
- Lacey, E. P. (1998). What is an adaptive environmentally induced parental effect?. In *Maternal Effects as Adaptations* (Mousseau, T.A. & Fox, C.W., eds), pp. 54–67. New York, NY: Oxford University Press.
- Lawrence, C., Ebersole, J. P. & Kesseli, R. V. (2008). Rapid growth and out-crossing promote female development in zebrafish (*Danio rerio*). Environmental Biology of Fishes 81, 239–246. doi: 10.1007/s10641-007-9195-8
- Leblanc, C. A. L., Benhaim, D., Hansen, B. R., Kristjansson, B. K. & Skulason, S. (2011). The importance of egg size and social effects of behaviour of Arctic charr juveniles. *Ethology* **117**, 664–674. doi: 10.1111/j.1439-0310.2011.01920.x
- Lee, J. S. F. & Berejikian, B. A. (2008). Effects of the rearing environment on average behaviour and behavioural variation in steelhead. *Journal of Fish Biology* **72**, 1736–1749. doi: 10.1111/j.1095-8649.2008.01848.x
- Lee, W.S., Monaghen, P. & Metcalfe, N.B. (2013*a*). The pattern of early growth trajectories affects adult breeding performance. *Ecology* **93**, 902–912. Available at http://dx.doi.org/10.1890/11-0890.1/
- Lee, W. S., Monaghen, P. & Metcalfe, N. B. (2013b). Environmental demonstration of the growth rate lifespan trade-off. *Proceedings of the Royal Society B* **280**, 20122370. doi: 10.1098/rspb.2012.2370
- Leips, J., Richardson, J. M. L., Rodd, F. H. & Travis, J. (2009). Adaptive maternal adjustments of offspring size in response to conspecific density in two populations of the least killifish, *Heterandria formosa*. *Evolution* **63**, 1341–1347. doi: 10.1111/j.1558-5646.2009.00631.x
- Lindsey, C. C. (1988). Factors controlling meristic variation. In *Fish Physiology*, Vol. 11 (Hoar, W. S. & Randall, D. J., eds), pp. 197–274. New York, NY: Elsevier.
- Lindström, J. (1999). Early development and fitness in birds and mammals. *Trends in Ecology and Evolution* **14**, 343–348.
- Lobon-Cervia, J., Utrilla, C., Rincón, P. & Amezcua, F. (1997). Environmentally induced spatio-temporal variations in the fecundity of brown trout *Salmo trutta* L.: trade-offs between egg size and number. *Freshwater Biology* **38**, 277–288. doi: 10.1046/j.1365-2427.1997.00217.x
- Lönnstedt, O. M., McCormick, M. I., Meekan, M. G., Ferrari, M. C. O. & Chivers, D. P. (2012). Learn and live: predator experience and feeding history determines prey behaviour and survival. *Proceedings of the Royal Society B* 279, 2091–2098. doi: 10.1098/rspb.2011.2516
- Lozano, C. A., Gjerde, B., Ødegård, J. & Bentsen, H. B. (2013). Heritability estimates for male proportion in the GIFT Nile tilapia (*Oreochromis niloticus* L.). *Aquaculture* **372–375**, 137–148. doi: 10.1016/j.aquaculture.2012.10.029
- Luckenbach, J. A., Godwin, J., Daniels, H. V. & Borski, R. J. (2003). Gonadal differentiation and effects of temperature on sex determination in southern flounder (*Paralichthys lethostigma*). *Aquaculture* **216**, 315–327. doi: 10.1016/S0044-8486(02)00407-6
- Lummaa, V. & Clutton-Brock, T. (2002). Early development, survival and reproduction in humans. *Trends in Ecology and Evolution* **17,** 141–147. doi: 10.1016/S0169-5347(01) 02414-4
- Madsen, T. & Shine, R. (2000). Silver spoons and snake body sizes: prey availability early in life influences long-term growth rates of free-ranging pythons. *Journal of Animal Ecology* **69**, 952–958.
- Magerhans, A., Muller-Belecke, A. & Horstgen-Schwark, S. (2009). Effect of rearing temperatures post hatching on sex ratios of rainbow trout (*Oncorhynchus mykiss*) populations. *Aquaculture* **294**, 25–29. doi: 10.1016/j.aquaculture.2009.05.001
- Malekpour, M. (2007). Effects of attachment on early and later development. *British Journal of Developmental Disabilities* **53**, 81–95.
- Marchetti, M. P. & Nevitt, G. A. (2003). Effects of hatchery rearing on brain structures of rainbow trout, *Oncorhynchus mykiss*. *Environmental Biology of Fishes* **66**, 9–14. doi: 10.1023/A:1023269221678

- Martell, D. J., Kieffer, J. D. & Tripple, E. A. (2005). Effects of temperature during early life history on embryonic and larval development and growth in haddock. *Journal of Fish Biology* **66**, 1558–1575. doi: 10.1111/j.0022-1112.2005.00699.x
- Martin, T. E., Ton, R. & Niklison, A. (2013). Intrinsic vs. extrinsic influences on life history expression: metabolism and parentally induced temperature influences on embryo development rate. *Ecology Letters* 16, 738–745. doi: 10.1111/ele.12103
- Matouk, C. C. & Marsden, P. A. (2008). Epigenetic regulation of vascular endothelial gene expression. *Circulation Research* 102, 873–887. doi: 10.1161/CIRCRE-SAHA.107.171025
- Mayer, I., Meager, J., Skjæraasen, J. E., Rodewald, P., Sverdrup, G. & Fernö, A. (2011). Domestication causes rapid changes in heart and brain morphology in Atlantic cod (*Gadus morhua*). Environmental Biology of Fishes 92, 181–186. doi: 10.1007/s10641-011-9831-1
- McAdam, A. G., Boutin, S., Réale, D. & Berteaux, D. (2002). Maternal effects and the potential for evolution in a natural population of animals. *Evolution* **56**, 846–851. doi: 10.1111/j.0014-3820.2002.tb01396.x
- McCormick, M. I. (1998). Behaviorally induced maternal stress in a fish influences progeny quality by a hormonal mechanism. *Ecology* **79**, 1873–1883.
- McCormick, C. M. & Green, M. R. (2013). From the stressed adolescent to the anxious and depressed adult: investigations in rodent models. *Neuroscience* **249**, 242–257. doi: 10.1016/j.neuroscience2012.08.063
- McGhee, K. E., Pintor, L. M., Suhr, E. L. & Bell, A. M. (2012). Maternal exposure to predation risk decreases offspring antipredator behaviour and survival in threespined stickleback. *Functional Ecology* **26**, 932–940. doi: 10.1111/j.1365-2435.2012.02008.x
- McGowan, R. A. & Martin, C. C. (1997). DNA methylation and genome imprinting in zebrafish, *Danio rerio*: some evolutionary ramifications. *Biochemistry and Cell Biology* **75**, 499–506.
- Mesoudi, A., Blanchet, S., Charmantier, A., Danchin, E., Fogarty, L., Jablonka, E., Laland, K. N., Morgan, T. H., Müller, G. B., Odling-Smee, F. J. & Pujol, B. (2013). Is non-genetic inheritance just a proximate mechanism? A corroboration of the extended evolutionary synthesis. *Biological Theory* 7, 189–195. doi: 10.1007/s13752-013-0091-5
- Metcalfe, N. B. & Monaghan, P. (2003). Growth versus lifespan: perspectives from evolutionary ecology. *Experimental Gerontology* **38**, 935–940.
- Mirbahai, L., Williams, T., Zhan, H., Gong, Z. & Chipman, J. K. (2011). Comprehensive profiling of zebrafish hepatic proximal promoter CpG island methylation and its modification during chemical carcinogenesis. *BMC Genomics* 12, 3.
- Montalvo, C., Villar, A. V., Merino, D., García, R., Ares, M., Llano, M., Cobo, M., Hurlé, M. A. & Nistal, J. F. (2012*a*). Androgens contribute to sex differences in myocardial remodelling under pressure overload by a mechanism involving TGF-*β*. *PLoS One* **7**, e35635. doi: 10.1371/journal.pone.0035635
- Montalvo, A. J., Faulk, C. K. & Holt, G. J. (2012b). Sex determination in southern flounder, *Paralichthys lethostigma*, from the Texas Gulf Coast. *Journal of Experimental Marine Biology and Ecology* **432**, 186–190. doi: 10.1016/j.jembe.2012.07.017
- Moran, P. & Pérez-Figueroa, A. (2011). Methylation changes associated with early maturation stages in the Atlantic salmon. *BMC Genetics* 2011 12, 86. doi: 10.1186/1471-2156-12-86
- Morita, K., Yamamoto, S., Takashima, Y., Matsuishi, T., Kanno, Y. & Nishimura, K. (1999). Effect of maternal growth history on egg number and size in wild white-spotted char (Salvelinus leucomaenis). Canadian Journal of Fisheries and Aquatic Sciences 56, 1585–1589. doi: 10.1139/cjfas-56-9-1585
- Morita, K., Tamate, T., Sugimoto, Y., Tago, Y., Watanabe, T., Konaka, H., Sato, M., Miyauchi, Y., Ohkuma, K. & Nagasawa, T. (2009). Latitudinal variation in egg size and number in anadromous masu salmon *Oncorhynchus masou*. *Journal of Fish Biology* **74**, 699–705. doi: 10.1111/j.1095-8649.2008.02150.x
- Morrongiello, J. R., Bond, N. R., Crook, D. A. & Wong, B. B. M. (2012). Spatial variation in egg size and egg number reflects trade-offs and bet-hedging in a freshwater fish. *Journal of Animal Ecology* **81**, 806–817. doi: 10.1111/j.1365-2656.2012.01961.x
- Mousseau, T. A. & Fox, C. W. (1998). The adaptive significance of maternal effects moms do more than just make babies. *Trends in Ecology and Evolution* **13**, 403–407.

- Näslund, J., Aarestrup, K., Thomassen, S. T. & Johnsson, J. (2012). Early enrichment effects on brain development in hatchery-reared Atlantic salmon (*Salmo salar*): no evidence for a critical period. *Canadian Journal of Fisheries and Aquatic Sciences* 69, 1481–1490. doi: 10.1139/f2012-074
- Navarro-Martin, L., Vinas, J., Ribas, L., Diaz, N., Gutierrez, A., Di Croce, L. & Piferrer, F. (2011). DNA methylation of the gonadal aromatase (cyp 19a) promotor is involved in temperature-dependent sex ratio shifts in the European sea bass. *PLoS Genetics* 7, e1002447. doi: 10.1371/journal.pgen.1002447
- Nelson, E. D. & Monteggia, L. M. (2011). Epigenetics in the mature mammalian brain: effects on behavior and synaptic transmission. *Neurobiology of Learning and Memory* **96**, 53–60. doi: 10.1016/j.nlm.2011.02.015
- Olsson, I. C., Greenberg, L. & Bergman, E. (2006). Environmentally induced migration: the importance of food. *Ecology Letters* **9**, 645–651. doi: 10.1111/j.1461-0248.2006.00909.x
- Omoto, N., Koya, Y., Chin, B., Yamashita, Y., Nakagawa, M. & Noda, T. (2010). Gonadal sex differentiation and effect of rearing temperature on sex ratio in black rockfish (*Sebastes schlegeli*). *Ichthyological Research* **57**, 133–138. doi: 10.1007/s10228-009-0137-7
- Ospina-Álvarez, N. & Piferrer, F. (2008). Temperature-dependent sex determination in fish revisited: prevalence, a single sex ratio response pattern, and possible effects of climate change. *PLoS One* **3**, e2837. doi: 10.1371/journal.pone.0002837
- Patino, R., Davis, K. B., Schoore, J. E., Uguz, C., Strüssmann, C. A., Parker, N. C., Simco, B. A. & Goudle, C. A. (1996). Sex differentiation of channel catfish gonads: normal development and effects of temperature. *Journal of Experimental Zoology* **276**, 209–218. doi: 10.1002/(SICI)1097-010X(19961015)276:3<209::AID-JEZ5>3.0.CO;2-R
- Paull, G. C., Filby, A. L. & Tyler, C. R. (2009). Growth rate during early life affects sexual differentiation in roach (*Rutilus rutilus*). Environmental Biology of Fishes **85**, 277–284. doi: 10.1007/s10641-009-9492-5
- Piché, J., Hutchings, J. A. & Blanchard, W. (2008). Genetic variation in threshold reaction norms for alternative reproductive tactics in male Atlantic salmon, *Salmo salar. Proceedings of* the Royal Society B 275, 1571–1575. doi: 10.1098/rspb.2008.0251
- Pörtner, H. O. (2010). Oxygen- and capacity-limitation of thermal tolerance: a matrix for integrating climate-related stressor effects in marine ecosystems. *Journal of Experimental Biology* **213**, 881–893. doi: 10.1242/jeb.037523
- Putman, N. F., Scanlan, M. M., Bilman, E. J., O'Neil, J. P., Couture, R. B., Quinn, T. P., Lohman, K. J. & Noakes, D. L. G. (2014). An inherited magnetic map guides ocean navigation in juvenile Pacific salmon. *Current Biology* **24**, 446–450. doi: 10.1016/j.cub.2014.01.017
- Quinn, T. P. (1980). Evidence for celestial magnetic compass orientation in lake migrating sockeys salmon fry. *Journal of Comparative Physiology* **137**, 243–248.
- Quinn, T. P., Vøllestad, L. A., Peterson, J. & Gallucci, V. (2004). Influences of freshwater and marine growth on the egg size-egg number trade off in coho and Chinook salmon. *Transactions of the American Fisheries Society* **133**, 55–65. doi: 10.1577/T03-032
- Quinn, T. P., Seamons, T. R., Vøllestad, L. A. & Duffy, E. (2011). Effects of growth and reproductive history on the egg size fecundity trade-off in steelhead. *Transactions of the American Fisheries Society* **140**, 45–51. doi: 10.1080/00028487.2010.550244
- Ramsahoye, B. H., Biniszkiewicz, D., Lyko, F., Clark, V., Bird, A. P. & Jaenisch, R. (2000). Non-CpG methylation is prevalent in embryonic stem cells and may be mediated by DNA methyltransferase 3a. *Proceedings of the National Academy of Sciences of the United States of America* 97, 5237–5242. doi: 10.1073/pnas.97.10.5237
- Räsänen, K. & Kruuk, L. E. B. (2007). Maternal effects and evolution at ecological time-scales. Functional Ecology 21, 408–421. doi: 10.1111/j.1365-2435.2007.01246.x
- Réale, D., Garant, D., Humphries, M. M., Bergeron, P., Careau, V. & Montiglio, P. O. (2010). Personality and the emergence of the pace-of-life syndrome concept at the population level. *Philosophical Transactions of the Royal Society B* 365, 4051–4063. doi: 10.1098/rstb.2010.0208
- Reebs, S. G. (1996). Time-place learning in golden shiners (Pisces: Cyprinidae). *Behavioural Processes* **36**, 253–262. doi: 10.1016/0376-6357(96)88023-5
- Reznick, D. (1991). Maternal effects in fish life histories. In *The Unity of Evolutionary Biology* (Dudley, E. C., ed), pp. 780–793. Portland, OR: Dioscorides Press.

- Richards, C. L., Bossdorf, O. & Pigluicci, M. (2010). What role does heritable epigenetic variation play in phenotypic evolution? *BioScience* **60**, 232–237. doi: 10.1525/bio.2010.60.3.9
- Riggs, A. D. (1975). X inactivation, differentiation, and DNA methylation. *Cytogenetics and Cell Genetics* **14**, 9–25.
- Roach, D. A. & Wulff, R. (1987). Maternal effects in plants: evidence and ecological and evolutionary significance. *Annual Review of Ecology and Systematics* **18**, 209–235.
- Roberts, S. B. & Gavery, M. R. (2012). Is there a relationship between DNA methylation and phenotypic plasticity in invertebrates? *Frontiers in Physiology* **2**, 1–5. doi: 10.1016/j.cbd.2011.12.003
- Roff, D. A. (1992). *The Evolution of Life Histories: Theory and Analysis*. New York, NY: Chapman & Hall.
- Römer, U. & Beisenherz, W. (1996). Environmental determination of sex in *Apistogramma* (Cichlidae) and two other freshwater fishes (Teleostei). *Journal of Fish Biology* **48**, 714–725. doi: 10.1111/j.1095-8649.1996.tb01467.x
- Rossiter, M. C. (1996). Incidence and consequences of inherited environmentally induced parental effects. *Annual Review of Ecology and Systematics* **27**, 451–476. doi: 10.1146/annurev.ecolsys.27.1.451
- Rougeot, C., Prignon, C., Kengne, C. V. N. & Melard, C. (2008). Effect of high temperature during embryogenesis on the sex differentiation process in the Nile tilapia, *Oreochromis niloticus*. *Aquaculture* **276**, 205–208. doi: 10.1016/j.aquaculture.2008.02.001
- Rubin, D. A. (1985). Effect of pH on sex ratio in cichlids and a poeciliid (Teleostei). *Copeia* **1985**, 233–235.
- Rungruangsak-Torrissen, K. & Male, R. (2000). Trypsin isozymes: development, digestion and structure. In *Seafood Enzymes: Utilization and Influence on Postharvest Seafood Quality* (Haard, N. F. & Simpson, B. K., eds), pp. 215–269. New York, NY: Marcel Dekker, Inc.
- Rungruangsak-Torrisen, K., Moss, R., Andresen, L.H., Berg, A. & Waagbø, R. (2006). Different expressions of trypsin and chymotrypsin in relation to growth in Atlantic salmon (*Salmo salar L.*). Fish Physiology and Biochemistry **32**, 7–23. doi: 10.1007/s10695-005-0630-5
- Sainz, J.C., García-Carreño, F.L., Córdova-Murueta, J.H. & Cruz-Hernández, P. (2005). Whiteleg shrimp (*Litopenaeus vannamei*, Boone, 1931) isotrypsins: their genotype and modulation. *Journal of Experimental Marine Biology and Ecology* 326, 105–113. doi: 10.1016/j.jembe.2005.05.021
- Salinas, S. & Munch, S. B. (2012). Thermal legacies: transgenerational effects of temperature on growth in a vertebrate. *Ecology Letters* **15**, 159–163. doi: 10.1111/j.1461-0248. 2011.01721.x
- Salvanes, A. G. V. & Braithwaite, V. A. (2005). Exposure to variable spatial information in the early rearing environment generates asymmetries in social interactions in cod (*Gadus morhua*). Behavioural Ecology and Sociobiology 59, 250–257. doi: 10.1007/s00265-005-0031-x
- Salvanes, A. G. V., Moberg, O., Ebbeson, L.O.E., Nilsen, T.O., Jensen, K.H. & Braithwaite, V. A. (2013). Environmental enrichment promotes neural plasticity and cognitive ability in fish. *Proceedings of the Royal Society B* 280, 3113. Available at http://dx.doi.org/10.1098/rspb.2013.1331/
- Sandra, G. É. & Norma, M. M. (2010). Sexual determination and differentiation in teleost fish. *Reviews in Fish and Fisheries* **20**, 101–121. doi: 10.1007/s11160-009-9123-4
- Scheiner, S. M. & Holt, D. H. (2012). The genetics of phenotypic plasticity. X. Variation versus uncertainty. *Ecology and Evolution* **2,** 751–762. doi: 10.1002/ece3.217
- Schlichting, C. D. & Pigliucci, M. (1998). *Phenotypic Evolution: A Reaction Norm Perspective*. Sunderland, MA: Sinauer Associates.
- Schultz, E. T. (2008). A sex difference in seasonal timing of birth in a livebearing fish. *Copeia* **2008**, 673–679. doi: 10.1643/CE-07-168
- Scott, G. R. & Johnston, I. A. (2012). Temperature during embryonic development has persistent effects on thermal acclimation capacity of zebrafish. *Proceedings of the National Academy of Sciences of the United States of America* 109, 14247–14252. doi: 10.1073/pnas.1205012109

- Segers, F. H. I. D. & Taborsky, B. (2011). Egg size and food abundance interactively affect juvenile growth and behaviour. *Functional Ecology* **25**, 166–176. doi: 10.1111/j.1365-2435.2010.01790.x
- Segers, F. H. İ. D. & Taborsky, B. (2012). Juvenile exposure to predator cues induces a larger egg size in fish. *Proceedings of the Royal Society B* **22**, 1241–1248. doi: 10.1098/repb2011.1290
- Segers, F. H. I. D., Berishvili, G. & Taborsky, B. (2012). Egg size-dependent expression of growth hormone receptor accompanies compensatory growth in fish. *Proceedings of the Royal Society B* 279, 592–600. doi: 10.1098/rspb.2011.1104
- Selim, K. M., Shinomiya, A., Otake, H., Hamaguchi, S. & Sakaizumi, M. (2009). Effects of high temperature on sex differentiation and germ cell production in medaka, *Oryzias latipes*. *Aquaculture* **289**, 340–349. doi: 10.1016/j.aquaculture.2008.12.019
- Senior, A. M., Nat, L. J. & Nakagawa, S. (2012). The fitness consequences of environmental sex reversal in fish: a quantitative review. *Biological Reviews* 87, 900–911. doi: 10.1111/j.1469-185X.2012.00230.x
- Sfakianakis, D. G., Leris, J., Laggis, A. & Kentouri, M. (2011). The effect of rearing temperature on body shape and meristic characters in zebrafish (*Danio rerio*) juveniles. *Environmental Biology of Fishes* **92**, 197–205. doi: 10.1007/s10641-011-9833-z
- Siegfried, K. R. (2010). In search of determinants: gene expression during gonadal sex differentiation. *Journal of Fish Biology* **76**, 1879–1902. doi: 10.1111/j.1095-8649.2010.02594.x
- Silverstein, J. T., Shimma, H. & Ogata, H. (1997). Early maturity in amago salmon (*Oncorhynchus masu ishikawai*): an association with energy storage. *Canadian Journal of Fisheries and Aquatic Sciences* **54**, 444–451. doi: 10.1139/f96-284
- Simonet, N.G., Reyes, M., Nardocci, G., Molina, A. & Alvarez, M. (2013). Epigenetic regulation of the ribosomal cistron seasonally modulates enrichment of H2A.Z and H2A.Zub in response to different environmental inputs in carp (*Cyprinus carpio*). *Epigenetics and Chromatin* **6**, 22. doi: 10.1186/1756-8935-6-22
- Singh, K., Molenaar, A. J., Swanson, K. M., Gudex, B., Arias, A. J., Erdman, R. A. & Stelwagen, K. (2012). Epigenetics: a possible role in acute and transgenerational regulation of dairy cow milk production. *Animal* **6**, 375–381. doi: 10.1017/S1751731111002564
- Stamps, J. & Groothuis, T. G. G. (2010). The development of animal personality: relevance, concepts and perspectives. *Biological Reviews* **85**, 301–325. doi: 10.1111/j.1469-185X. 2009.00103.x
- Stearns, S. C. (1992). *The Evolution of Life Histories*. New York, NY: Oxford University Press. Steinbacher, P., Marschallinger, J., Obermayer, A., Neuhofer, A., Sanger, A. M. & Stoiber, W. (2011). Temperature-dependent modification of muscle precursor cell behaviour is an underlaying reason for lasting effects on muscle cellularity and body growth of teleost fish. *Journal of Experimental Biology* **214**, 1791–1801. doi: 10.1242/jeb.050096
- Stickland, N. C., White, R. N., Mescall, P. E., Crook, A. R. & Thorpe, J. E. (1988). The effect of temperature on myogenesis in embryonic development of the Atlantic salmon (*Salmo salar L.*). *Anatomy and Embryology* **178**, 253–257.
- Strand, D. A., Utne-Palm, A. C., Jakobsen, P. J., Braithwaite, V. A., Jensen, K. H. & Salvanes, A. G. V. (2010). Enrichment promotes learning in fish. *Marine Ecology Progress Series* **412**, 273–282. doi: 10.3354/meps08682
- Stratman, A. & Taborsky, B. (2014). Antipredator defences of young are independently determined by genetic inheritance, maternal effects and own early experience in mouthbrooding cichlids. *Functional Ecology*. doi: 10.1111/1365-2435.12224
- Stuart-Fox, D. & Moussalli, A. (2008). Selection for social signalling drives the evolution of chameleon colour change. *PLoS Biology* **6**, e25. doi: 10.1371/journal.pbio.0060025
- Sundström, L. F., Petersson, E., Höjesjö, J., Johnsson, J. I. & Järvi, T. (2004). Hatchery selection promotes boldness in newly hatched brown trout (*Salmo trutta*): implications for dominance. *Behavioral Ecology* 15, 192–198. doi: 10.1093/beheco/arg089
- Tåning, A. V. (1952). Experimental study of meristic characters in fishes. *Biological Reviews* **27**, 169–193.
- Taborsky, B. (2006). Mothers determine offspring size in response to own juvenile growth conditions. *Biology Letters* **2**, 225–228. doi: 10.1098/rsbl.2005.0422

- Taborsky, B., Skubic, E. & Bruintjes, R. (2007). Mothers adjust egg size to helper number in a cooperatively breeding cichlid. *Behavioral Ecology* **18**, 652–657. doi: 10.1093/beheco/arm026
- Taborsky, B., Tschirren, L., Meunier, C. & Aubin-Horth, N. (2013). Stable reprogramming of brain transcription profiles by the early social environment in a cooperatively breeding fish. *Proceedings of the Royal Society B* 280, 20122605. doi: 10.1098/rspb.2012.2605
- Tamate, T. & Maekawa, K. (2000). Interpopulation variation in reproductive traits of female masu salmon, *Oncorhynchus masou*. *Oikos* **90,** 209–218. doi: 10.1034/j.1600-0706. 2000.900201.x
- Tessema, M., Müller-Belecke, A. & Hörstgen-Schwark, G. (2006). Effects of rearing temperatures on the sex ratios of *Oreochromis niloticus* populations. *Aquaculture* **258**, 270–277. doi: 10.1016/j.aquaculture.2006.04.041
- Thorpe, J. E. (1986). Age at first maturity on Atlantic salmon, *Salmo salar*: freshwater period influences and conflicts with smolting. *Canadian Special Publication of Fisheries and Aquatic Sciences* **89**, 7–14.
- Tomizawa, S., Kobayashi, H., Watanabe, T., Andrews, S., Hata, K., Kelsey, G. & Sasaki, H. (2011). Dynamic stage-specific changes in imprinted differentially methylated regions during early mammalian development and prevalence of non-CpG methylation in oocytes. *Development* 138, 811–820. doi: 10.1242/dev.061416
- Uusi-Heikkila, S., Kuparinen, A., Wolter, C., Meinelt, T. & Arlinghaus, R. (2012). Paternal body size affects reproductive success in laboratory-helt zebrafish (*Danio rerio*). *Environmental Biology of Fishes* **93**, 461–474. doi: 10.1007/s10641-011-9937-5
- Valenzuela, N., Adams, D. C. & Janzen, F. J. (2003). Pattern does not equal process: exactly when is sex environmentally determined? *American Naturalist* **161**, 676–683. doi: 10.1086/368292
- Verzijden, M. N. & Cate, C. (2007). Early learning influences assortative mating in Lake Victoria cichlid fish. *Biology Letters* **3**, 134–136. doi: 10.1098/rsbl.2006.0601
- Villamizar, N., Ribas, L., Piferrer, F., Vera, L. M. & Sánchez-Vázquez, F. J. (2012). Impact of daily thermocycles on hatching rhythms, larval performance and sex differentiation of zebrafish. *PLoS One* 7, e52153. doi: 10.1371/journal.pone.0052153
- Vøllestad, L. A., Peterson, J. & Quinn, T. P. (2004). Effects of freshwater and marine growth rates on early maturity in male coho and Chinook salmon. *Transactions of the American Fisheries Society* 133, 495–503. doi: 10.1577/T03-033.1
- Walsh, M. R. & Reznick, D. N. (2011). Experimentally induced life-history evolution in a killifish in response to the introduction of guppies. *Evolution* **65**, 1021–1036.
- Warburton, K. (2003). Learning of foraging skills by fish. *Fish and Fisheries* **4**, 203–215. doi: 10.1046/j.1467-2979.2003.00125.x
- Warner, D.A. & Shine, R. (2008). The adaptive significance of temperature-dependent sex determination in a reptile. *Nature* **451**, 566–568. doi: 10.1038/nature06519
- Warner, D. A., Jorgensen, C. F. & Janzen, F. J. (2010). Maternal and abiotic effects on egg mortality and hatchling size of turtles: temporal variation in selection over seven years. *Functional Ecology* **24**, 857–866. doi: 10.1111/j.1365-2435.2010.01714.x
- Wedekin, C., Evanno, G., Szekely, T., Pompini, M., Darbellay, O. & Guthruf, J. (2013). Persistent unequal sex ratio in a population of grayling (Salmonidae) and possible role of temperature increase. *Conservation Biology* 27, 229–234. doi: 10.1111/j.1523-1739. 2012.01909.x
- Werner, E. E. (1977). Species packing and niche complementarity in three sunfishes. *American Naturalist* **111**, 553–578.
- Werner, E. E. & Gilliam, J. F. (1984). The ontogenetic niche and species interactions in size-structured populations. *Annual Review of Ecology and Systematics* **15**, 393–425.
- Wimberger, P. H. (1992). Plasticity of fish body shape. The effects of diet, development, family and age in two species of *Geophagus* (Pisces; Cichlidae). *Biological Journal of the Linnean Society* **45**, 197–218. doi: 10.1111/j.1095-8312.1992.tb00640.x
- Wootton, R. J. (1998). *Ecology of Teleost Fishes*, 2nd edn. Dordrecht: Kluwer Academic Publisher.
- Wright, S. (1969). Evolution and the Genetics of Populations, Vol. 2. Chicago, IL: University of Chicago Press.

- Wysujack, K., Greenberg, L. A., Bergman, E. & Olsson, I. C. (2009). The role of the environment in partial migration: food availability affects the adoption of a migratory tactic in brown trout *Salmo trutta*. *Ecology of Freshwater Fish* **18**, 52–59. doi: 10.1111/j.1600-0633.2008.00322.x
- Yamaguchi, T., Yoshinaga, N., Yazawa, T., Gen, T. & Kitano, T. (2010). Cortisol is involved in temperature-dependent sex determination in the Japanese flounder. *Endocrinology* **151**, 3900–3908. doi: 10.1210/en.2010-0228
- Zhang, Y., Zhang, S., Liu, Z., Zhang, L. & Zhang, W. (2013). Epigenetic modifications during sex change repress gonadotropin stimulation of *Cyp19a1a* in a teleost ricefield eel (*Monopterus albus*). *Endocrinology* **154**, 2881–2890. doi: 10.1210/en.2012-2220